

Validation Workshop

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<http://www.cstl.nist.gov/biotech/strbase/validation.htm>



Pennsylvania State Police
DNA Laboratory Training Workshop
Greensburg, PA
June 5, 2007



Proposed Agenda for PSP Training

- Introductions – Presenter and Participants
 - **Validation Workshop**
- Lunch**
- *Answer questions*
 - Cover other topics (depending on time)
 - Mixture Interpretation
 - qPCR and Low-Copy Number (LCN) DNA Testing

My Goal is to Answer YOUR Questions – So Please Ask Them...

NIST and NIJ Disclaimer

Funding: Interagency Agreement 2003-IJ-R-029
between the National Institute of Justice and NIST
Office of Law Enforcement Standards

Points of view are mine and do not necessarily represent the official position or policies of the US Department of Justice or the National Institute of Standards and Technology.

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Introductions

My Background

- **My mom grew up in Pittsburgh and my parents met at Penn State so I have Pennsylvania roots...**
- PhD (**Analytical Chemistry**) from University of Virginia (Aug 1995)
- Research conducted at FBI Academy under Bruce McCord doing CE for STR typing
- NIST Postdoc – developed STRBase website
- GeneTrace Systems – **private sector experience validating assays**
- NIST Human Identity Project Leader since 1999
- Invited guest to SWGDAM since 2000
- **Member of SWGDAM Validation Subcommittee**
- Served on WTC KADAP and helped evaluate and validate new miniSTR, mtDNA, and SNP assays
- Author of *Forensic DNA Typing: Biology, Technology, and Genetics of STR Markers* (2nd Edition)
- Married with 6 children – I have "validated" that they are mine using STR typing...

NIST History and Mission

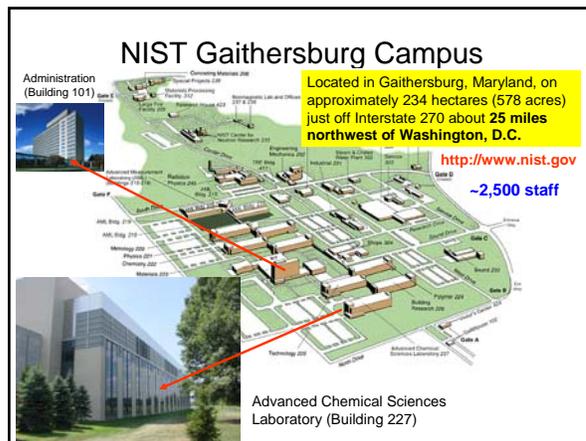
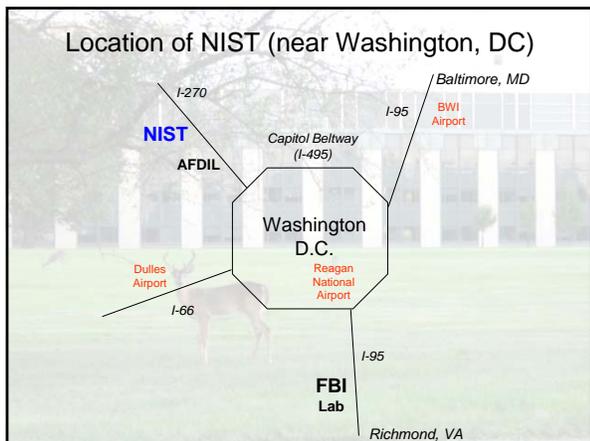
- National Institute of Standards and Technology (NIST) was created in 1901 as the National Bureau of Standards (NBS). The name was changed to NIST in 1988.
- NIST is **part of the U.S. Department of Commerce with a mission to develop and promote measurement, standards, and technology to enhance productivity, facilitate trade, and improve the quality of life.**
- NIST supplies over 1,300 Standard Reference Materials (SRMs) for industry, academia, and government **use in calibration of measurements.**
- **NIST defines time for the U.S.**



\$573 for 3 jars



DNA typing standard



NIST Human Identity Project Team

John Butler, Margaret Kline, Pete Vallone, Jan Redman, Amy Decker, Becky Hill, Dave Duerwer

All NIST publications and presentations available on STRBase:
<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

Human Identity Project Team
 Leading the Way In Forensic DNA...

- 26 publications from Jan-Dec 2006
- 45 presentations and 10 workshops to the community from Jan-Dec 2006

National Institute of Justice
 The Research, Development, and Evaluation Agency of the U.S. Department of Justice

Current Areas of NIST Effort with Forensic DNA

- **Standards** <http://www.cstl.nist.gov/biotech/strbase/>
 - Standard Reference Materials
 - Standard Information Resources (STRBase website)
 - Interlaboratory Studies
- **Technology**
 - Research programs in SNPs, miniSTRs, Y-STRs, mtDNA, qPCR
 - Assay and software development
- **Training Materials**
 - Review articles and workshops on STRs, CE, validation
 - PowerPoint and pdf files available for download

Seminars and Training Workshops to Individual Forensic DNA Laboratories

| | | | |
|--|--|-------------------------------|---|
| MYRIAD® Feb 3, 2005 | FBI LABORATORY May 19, 2005 | STATE POLICE June 8, 2005 | STATE POLICE June 13-14, 2005 |
| IOWA STATE UNIVERSITY Apr 27-28, 2006 | TEXAS STATE LABORATORY June 6, 2006 | BCA Aug 7, 2006 | STATE POLICE LABORATORY Nov 15, 2006 |
| BFS March 7, 2007 | STATE POLICE March 14, 2007 | STATE POLICE Apr 3-4, 2007 | STATE POLICE Apr 5, 2007 |
| STATE POLICE Dec 5-6, 2006 | STATE POLICE June 5, 2007 | STATE POLICE June 5, 2007 | STATE POLICE June 5, 2007 |

Training Workshops Planned

- **ISFG Meeting** (August 2007, Copenhagen, Denmark)
 - CE Fundamentals and Troubleshooting
 - Validation
- **SAFS Meeting** (September 2007, Atlanta, GA)
 - Mixture Interpretation
- **Int. Symposium on Human Identification (Promega) Meeting** (October 2007, Hollywood, CA)
 - Validation

Background of Participants...

- Name
- Experience (years) with DNA typing
- Something memorable about yourself
- What you hope to learn from this workshop



Background, Essentials and Importance of Validation

My Purpose in Teaching This Workshop

- I believe that many forensic laboratories, in an effort to be cautious, are taking too long to perform their validation studies and thereby delaying initiation of casework and contributing to backlogs in labs that are already overburdened
- Technology will continue to advance and thus validation of new methodologies will always be important in forensic DNA laboratories

There will always be something to “validate”...

Validation Workshop (Aug 24-26, 2005 at NFSTC)

<http://www.cstl.nist.gov/biotech/strbase/validation/validationworkshop.htm>

NIJ National Forensic Science Technology Center
President's DNA Initiative - Workshops

Validation Workshop

Robyn Ragsdale, PhD
Florida Department of Law Enforcement (FDLE)

John M. Butler, PhD
National Institute of Standards and Technology (NIST)

COURSE CONTENTS

Day #1

- Validation Overview (John)
- Introduction to DAB Standards (Robyn & John)
- Developmental Validation (John)

Day #2

- Inconsistency in Validation between Labs (John)
- Internal Validation (Robyn)
- Method Modifications and Performance Checks (Robyn)

Day #3

- Practical Exercises (Robyn)

Was filmed and is being made into a training DVD as part of the President's DNA Initiative...

Validation Workshop Outline

- Importance of Validation **Theoretical**
 - History of Forensic Validation Guidelines
 - SWGAM Revised Validation Guidelines**
 - Validation Philosophy & Resources
- BREAK**
- Developmental Validation **Practical**
 - Internal Validation and Examples
 - Documentation

Importance of Validation

Questions to Keep in Mind...

- Why is validation important?
- How does validation help with quality assurance within a laboratory?
- What are the general goals of analytical validation?
- How is method validation performed in other fields such as the pharmaceutical industry?
- How do accuracy, precision, sensitivity, stability, reproducibility, and robustness impact measurements?

What is **Validation** and Why Should It Be Done?

- Part of overall quality assurance program in a laboratory
- **We want the correct answer when collecting data...**
 - We want **analytical measurements made in one location to be consistent with those made elsewhere** (without this guarantee there is no way that a national DNA database can be successful).
- If we fail to get a result from a sample, we want to have confidence that the sample contains no DNA rather than there might have been something wrong with the detection method...

Want no false negatives...

Why is Method Validation Necessary?

- It is an important element of quality control.
- Validation helps provide assurance that a measurement will be reliable.
- In some fields, validation of methods is a regulatory requirement.
- ...
- The validation of methods is **good science**.

Roper, P., et al. (2001) *Applications of Reference Materials in Analytical Chemistry*. Royal Society of Chemistry, Cambridge, UK, pp. 107-108.

Definition of Validation

- **Validation** is confirmation by examination and provision of objective evidence that the particular requirements for a specified intended use are fulfilled.
- **Method validation** is the process of **establishing the performance characteristics and limitations of a method** and the identification of the influences which may change these characteristics and to what extent. It is also the process of verifying that a method is fit for purpose, i.e., for use for solving a particular analytical problem.

EURACHEM Guide (1998) *The Fitness for Purpose of Analytical Methods: A Laboratory Guide to Method Validation and Related Topics*; available at <http://www.eurachem.ul.pt/guides/valid.pdf>

More Validation Definitions

ISO 17025

5.4.5.1 Validation is the **confirmation by examination** and the provision of objective evidence that the particular requirements for a specific intended use are fulfilled

DAB Quality Assurance Standards for Forensic DNA Testing Laboratories

2 (ff) Validation is a **process by which a procedure is evaluated** to determine its efficacy and reliability for forensic casework analysis and includes:

To demonstrate that a method is suitable for its intended purpose...

Definitions

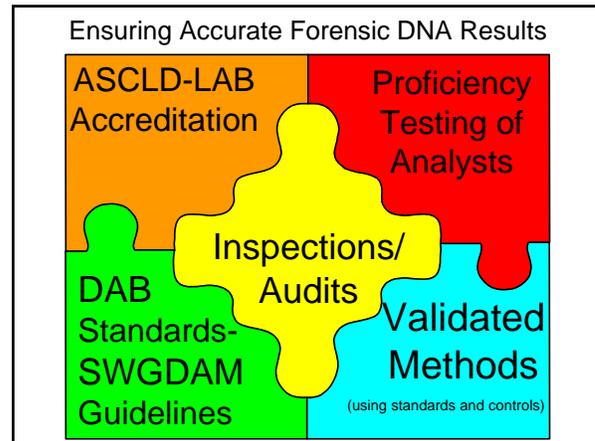
J.M. Butler (2005) *Forensic DNA Typing*, 2nd Edition, p. 389, 391

- **Quality assurance (QA)** – planned or systematic actions necessary to provide adequate confidence that a product or service will satisfy given requirements for quality
- **Quality control (QC)** – day-to-day operational techniques and activities used to fulfill requirements of quality
- **Validation** – the process of demonstrating that a laboratory procedure is **robust, reliable**, and **reproducible** in the hands of the personnel performing the test in that laboratory

Definitions

J.M. Butler (2005) *Forensic DNA Typing*, 2nd Edition, p. 391

- **Robust method** – successful results are obtained a high percentage of the time and few, if any, samples need to be repeated
- **Reliable method** – the obtained results are accurate and correctly reflect the sample being tested
- **Reproducible method** – the same or very similar results are obtained each time a sample is tested



Checks and Controls on DNA Results

| | |
|--------------------------------|--|
| Community | FBI DNA Advisory Board's Quality Assurance Standards (<i>also interlaboratory studies</i>) |
| Laboratory | ASCLD/LAB Accreditation and Audits |
| Analyst | Proficiency Tests & Continuing Education |
| Method/Instrument | Validation of Performance (<i>along with traceable standard sample</i>) |
| Protocol | Standard Operating Procedure is followed |
| Data Sets | Allelic ladders, positive and negative amplification controls, and reagent blanks are used |
| Individual Sample | Internal size standard present in every sample |
| Interpretation of Result | Second review by qualified analyst/supervisor |
| Court Presentation of Evidence | Defense attorneys and experts with power of discovery requests |

History of Forensic DNA Validation Guidelines

Timeline Regarding Forensic Validation Information

- 1989 *Castro case* – concern over quality in forensic DNA cases
- 1989 TWGDAM – focus on RFLP
- 1991 TWGDAM – updated with PCR info
- 1995 TWGDAM – updated with more PCR info
- **1998/99 DNA Advisory Board Standards**
- 2004 Revised SWGDAM Validation Guidelines

Brief Historical Overview

Profiles in DNA (Sept 1999) 3(2): 10-11

CURRENT EVENTS

The Evolution of Quality Standards for Forensic DNA Analyses in the United States

*By Special Agent Lawrence A. Presley, MS, MA
 Federal Bureau of Investigation Laboratory, Washington, DC
 lpresley@fbi.gov*

Quality problems in late 1980s with DNA testing
 TWGDAM established under FBI Lab sponsorship in 1988
 NRC I (1992) and NRC II (1996) issued reports recommending formal QA programs
DNA Identification Act of 1994 lead to formation of DNA Advisory Board (DAB)
 DAB Standards issued in Oct 1998 and Apr 1999
 When DAB was dissolved in 2000, SWGDAM assumed leadership role

DNA Identification Act (1994)

Public Law 103-322

42 § 14131. Quality assurance and proficiency testing standards

(a) Publication of quality assurance and proficiency testing standards

(1) (A) Not later than 180 days after September 13, 1994, the Director of the Federal Bureau of Investigation shall appoint an advisory board on DNA quality assurance methods from among nominations proposed by the head of the National Academy of Sciences and professional societies of crime laboratory officials.

(B) The advisory board shall include as members scientists from State, local, and private forensic laboratories, molecular geneticists and population geneticists not affiliated with a forensic laboratory, and a representative from the National Institute of Standards and Technology.

(C) **The advisory board shall develop, and if appropriate, periodically revise, recommended standards for quality assurance**, including standards for testing the proficiency of forensic laboratories, and forensic analysts, in conducting analyses of DNA.

DNA Advisory Board (DAB)

DNA Advisory Board (DAB) Members

- Joshua Lederberg (Rockefeller University) – chair 1995-1998
- Arthur Eisenberg (University of North Texas Health Science Center) – chair 1998-2000
- John Hicks (Alabama Department of Forensic Sciences)
- Shirley Abrahamson (Wisconsin State Supreme Court)
- Ranajit Chakraborty (University of Texas Health Science Center)
- Bruce Budowle (FBI Laboratory)
- Larry Presley (FBI Laboratory)
- Jack Ballantyne (Suffolk County Crime Lab)
- Jay Miller (FBI Laboratory)
- Dennis Reeder (National Institute of Standards and Technology)
- Margaret Kuo (Orange County Sheriff's Office)
- Bernard Devlin (Carnegie Mellon University)
- Marcia Eisenberg (Laboratory Corporation of America)
- Paul Ferrara (Virginia Division of Forensic Science)
- Terry Lober (Minnesota State DNA Lab)
- Dwight Adams, Randall Murch, Barry Brown (FBI Laboratory)
- David Coffman (Florida Department of Law Enforcement)
- Fred Bieber (Harvard Medical School)
- Mary Gibbons (Oakland Police Department)
- Eric Jungst (Case Western Reserve University)
- Susan Narveson (Phoenix Police Department)
- Mohammad Tahir (Indianapolis-Marion County Crime Lab)
- Dawn Herkenham (FBI Laboratory)

Existed from 1995-2000

DAB Standards

DNA Advisory Board
July 1998
(Part 1 of 2)

Text in red font from Quality Assurance Standards for Convicted Offender DNA Databasing Laboratories (April 1999)

<http://www.fbi.gov/hq/lab/fsc/backissu/july2000/codis2a.htm>

<http://www.fbi.gov/hq/lab/fsc/backissu/july2000/codis1a.htm>

Outline of DAB Standards

Quality Assurance Standards (QAS)

1. SCOPE
2. DEFINITIONS
3. QUALITY ASSURANCE PROGRAM
4. ORGANIZATION AND MANAGEMENT
5. PERSONNEL
6. FACILITIES
7. EVIDENCE (SAMPLE) CONTROL
- 8. VALIDATION**
9. ANALYTICAL PROCEDURES
10. EQUIPMENT CALIBRATION AND MAINTENANCE
11. REPORTS
12. REVIEW
13. PROFICIENCY TESTING
14. CORRECTIVE ACTION
15. AUDITS
16. SAFETY
17. SUBCONTRACTOR OF ANALYTICAL TESTING FOR WHICH VALIDATED PROCEDURES EXIST

Revised SWGDAM Validation Guidelines

(July 2004)

http://www.fbi.gov/hq/lab/fsc/backissu/july2004/standards/2004_03_standards02.htm

Forensic Science Communications July 2004 – Volume 6 – Number 3
Standards and Guidelines

Revised Validation Guidelines

Scientific Working Group on DNA Analysis Methods (SWGDM)

Introduction | Validation Considerations | Developmental Validation | Internal Validation | Material Modification | Performance Check | Definitions

Introduction

The validation section of the Guidelines for a Quality Assurance Program for DNA Analysis by the Technical Working Group on DNA Analysis Methods (*Crime Laboratory Digest* 1995:22(2):21-43) has been revised due to increased laboratory experience, the advent of new technologies, and the issuance of the Quality Assurance Standards for Forensic DNA Testing Laboratories by the Director of the FBI (Forensic Science Communications available: www.fbi.gov/hq/lab/fsc/backissu/july2000/codis2a.htm)

The document provides validation guidelines and definitions approved by SWGDAM July 10, 2003.

Validation Philosophy

When is Validation Needed?

- Before introduction of a new method into routine use
- Whenever the conditions change for which a method has been validated, e.g., instrument with different characteristics
- Whenever the method is changed, and the change is outside the original scope of the method

L. Huber (2001) Validation of Analytical Methods: Review and Strategy. Supplied by www.labcompliance.com

Costs/Benefits of Validation and Quality Assurance

Costs

- Direct
 - Test materials
 - Standards
 - Quality assurance equipment
 - Analysis of QA/QC samples
 - Quality assurance official
 - Committee Work
 - Interlab Studies
 - Travel to meetings

Benefits

- More efficient outputs
- Fewer replicates for same reliability
- Fewer do-overs
- Greater confidence of:
 - Staff
 - Laboratory
 - Customers

Table 26.2 in J.K. Taylor (1987) *Quality Assurance of Chemical Measurements*. Lewis Publishers: Chelsea, MI.

Some Purposes of Validation

- To accept an individual sample as a member of a population under study
- To admit samples to the measurement process
- To minimize later questions on sample authenticity
- To provide an opportunity for resampling when needed

Sample validation should be based on objective criteria to eliminate subjective decisions...

J.K. Taylor (1987) *Quality Assurance of Chemical Measurements*. Lewis Publishers: Chelsea, MI, p. 193

The VAM Principles

VAM = Valid Analytical Measurement

1. Analytical measurements should be made to satisfy an agreed requirement.
2. Analytical measurements should be made using methods and equipment that have been tested to ensure they are fit for their purpose.
3. **Staff making analytical measurements should be both qualified and competent to undertake the task.**
4. There should be a regular and independent assessment of the technical performance of a laboratory.
5. **Analytical measurements made in one location should be consistent with those made elsewhere.**
6. Organizations making analytical measurements should have well defined quality control and quality assurance procedures.

Roper P et al. (2001) *Applications of Reference Materials in Analytical Chemistry*. Royal Society of Chemistry: Cambridge UK, p. 2

Community Needs Training

- To better understand what validation entails and how it should be performed (why a particular data set is sufficient)
- Many labs already treat DNA as a "black box" and therefore simply want a "recipe" to follow
- People are currently driven by fear of auditors and courts rather than scientific reasoning
- Many different opinions exist and complete consensus is probably impossible

How do you validate a method?

- Decide on analytical requirements
 - Sensitivity, resolution, precision, etc.
- **Plan a suite of experiments**
- **Carry out experiments**
- Use data to assess fitness for purpose
- Produce a statement of validation
 - Scope of the method

Roper, P., et al. (2001) *Applications of Reference Materials in Analytical Chemistry*. Royal Society of Chemistry, Cambridge, UK, pp. 108-109.

Assumptions When Performing Validation

- The equipment on which the work is being done is broadly suited to the application. It is clean, well-maintained and **within calibration**.
- The staff carrying out the validation are **competent** in the type of work involved.
- There are **no unusual fluctuations in laboratory conditions** and there is no work being carried out in the immediate vicinity that is likely to cause interferences.
- The samples being used in the validation study are known to be **sufficiently stable**.

Roper, P., et al. (2001) *Applications of Reference Materials in Analytical Chemistry*. Royal Society of Chemistry, Cambridge, UK, pp. 110-111.

Tools of Method Validation

- Standard samples
 - positive controls
 - NIST SRMs
- Blanks
- Reference materials prepared in-house and spikes
- Existing samples
- Statistics
- **Common sense**

Roper, P., et al. (2001) *Applications of Reference Materials in Analytical Chemistry*. Royal Society of Chemistry, Cambridge, UK, p. 110.

Recent Articles I Have Written on Validation

Profiles in DNA (Promega Corporation), vol. 9(2), pp. 3-6

VALIDATION

http://www.promega.com/profiles/902/ProfileInDNA_902_03.pdf

Debunking Some Urban Legends Surrounding Validation Within the Forensic DNA Community

By John Butler
 National Institute of Standards and Technology, Gaithersburg, Maryland, USA

http://marketing.appliedbiosystems.com/images/forensic/volume8/PDFs_submitted/02A_CustomerCorner_Val_What_is_it.pdf

Applied Biosystems

Forensic News

January 2007 Customer Corner

Validation: What is it, Why Does it Matter, and How Should it Be Done?
 By John M. Butler, National Institute of Standards and Technology

Validation involves performing laboratory tests to verify that a particular instrument, software program, or measurement technique is working properly. These validation experiments typically examine precision, accuracy, and sensitivity, which all play a factor on the 3 R's of measurements: reliability, repeatability, and robustness.

Urban Legends of Validation...

Butler, J.M. (2006) *Profiles in DNA* vol. 9(2), pp. 3-6

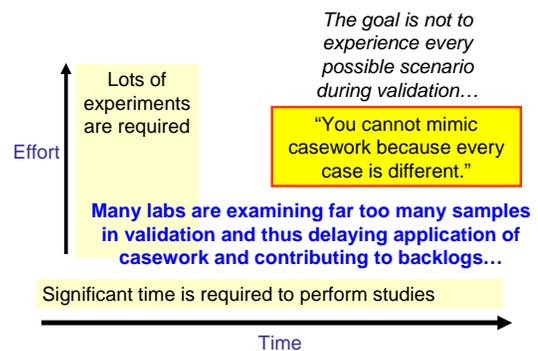
- #1: HUNDREDS OR THOUSANDS OF SAMPLES ARE REQUIRED TO FULLY VALIDATE AN INSTRUMENT OR METHOD
- #2: VALIDATION IS UNIFORMLY PERFORMED THROUGHOUT THE COMMUNITY
- #3: EACH COMPONENT OF A DNA TEST OR PROCESS MUST BE VALIDATED SEPARATELY
- #4: VALIDATION SHOULD SEEK TO UNDERSTAND EVERYTHING THAT COULD POTENTIALLY GO WRONG WITH AN INSTRUMENT OR TECHNIQUE
- #5: LEARNING THE TECHNIQUE AND TRAINING OTHER ANALYSTS ARE PART OF VALIDATION
- #6: VALIDATION IS BORING AND SHOULD BE PERFORMED BY SUMMER INTERNS SINCE IT IS BENEATH THE DIGNITY OF A QUALIFIED ANALYST
- #7: DOCUMENTING VALIDATION IS DIFFICULT AND SHOULD BE EXTENSIVE
- #8: ONCE A VALIDATION STUDY IS COMPLETED YOU NEVER HAVE TO REVISIT IT

My Philosophy towards Validation

Ask first: Does the new method improve your capability?

- **Concordance** – are the same typing results obtained with the new technique as with an older one?
- **Constant Monitoring** – check multiple allelic ladders in a batch against one another to confirm precision and consistent lab temperature
- **Common Sense** – are replicate tests repeatable?

Common Perceptions of Validation



Number of Samples Needed

Relationship between a sample and a population of data

Data collected in your lab as part of validation studies

"Sample" of Typical Data

How do you relate these two values?

Student's *t*-Test associates a sample to a population

All potential data that will be collected in the future in your lab

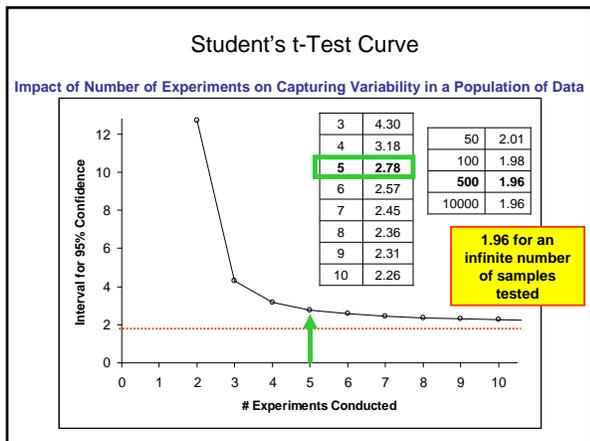
"Population" of All Data Obtained

Student's *t*-Tests

"Student" (real name: W. S. Gossett [1876-1937]) developed statistical methods to solve problems stemming from his employment in a brewery.

Student's *t*-test deals with the problems associated with inference based on "small" samples: the calculated mean (X_{avg}) and standard deviation (σ) may by chance deviate from the "real" mean and standard deviation (i.e., **what you'd measure if you had many more data items: a "large" sample**).

<http://www.physics.csbsju.edu/stats/t-test.html>



The Number "5" in Forensic Validation

NDIS Appendix B Expert System Validation Requirements

- At least **5 challenge events** must be observed for each issue (e.g., pullup, shoulders, spikes, tri-allelic patterns, mixtures, contamination, variant alleles)

Allele Frequency Tables

Butler et al. (2003)
JFS 48(4):908-911

Einum et al. (2004)
JFS 49(6): 1381-1385

Allele frequencies denoted with an asterisk (*) are below the 5/2N minimum allele threshold recommended by the National Research Council report (NRCII) The Evaluation of Forensic DNA Evidence published in 1996.

| Allele | Caucasian | |
|------------------------------|---------------|---------------|
| | N= 302 | N= 7,636 |
| 11 | 0.0017* | 0.0009 |
| 12 | 0.0017* | 0.0007 |
| 13 | -- | 0.0031 |
| 14 | 0.1027 | 0.1240 |
| Most common allele 15 | 0.2616 | 0.2690 |
| 15.2 | -- | -- |
| 16 | 0.2533 | 0.2430 |
| 17 | 0.2152 | 0.2000 |
| 18 | 0.15232 | 0.1460 |
| 19 | 0.01160 | 0.0125 |
| 20 | 0.0017* | 0.0001* |

Minimum Allele Frequency = **5/2N**

Want to sample at least 5 chromosomes to provide a somewhat reliable estimate of an allele's frequency in a population

What are the goals of validation studies involving a new STR typing kit?

- Stutter product amounts
Why?: aids in mixture interpretation guidelines (how often does laboratory call peaks below 15% of an adjacent allele?)
- Precision studies
Why?: aids in defining allele bin widths (how often does laboratory change the ± 0.5 bp from the mean?)
- Sensitivity studies
Why?: aids in detecting the minor component (how often does laboratory call peaks below 10% of the major component?)
- Multiplexing studies
Why?: aids in detecting the minor component (how often does laboratory call peaks below 10% of the major component?)
- Peak height ratio studies
Why?: aids in mixture interpretation guidelines (how often does your laboratory call peaks below a 60% heterozygote peak height ratio?)
- Primer sets
Why?: aids in mixture interpretation guidelines (how often does your laboratory call peaks below a 60% heterozygote peak height ratio?)

Too often validation experiments are performed but observations are not considered for implementation purposes

**FBI DNA Quality Assurance Audit
 Developmental Validation Scorecard**

Discussion
 Developmental validation must precede the introduction of a novel method. A novel methodology may include an existing technology or testing of a specific technology (e.g., medical testing, genetic analysis, or DNA analysis). Citations in peer-reviewed scientific journals and other sources of novel methodology should be available.

8.1.2

Validation Experiments Are Sometimes Driven by Fear of Auditors Rather than Good Science

8.2.3.1(FO-b)

FBI DNA Quality Assurance Audit Document
 Issue Date 07/04 (Rev. #6) 28

**Validation in Other Fields
 (Besides Forensic DNA Testing)**

**Pharmaceutical Industry and FDA Follows
 ICH Validation Documents**

- ICH (**I**nternational **C**onference on **H**armonization of Technical Requirements for Registration of Pharmaceuticals for Human Use)
 - <http://www.ich.org>
 - **Q2A: Text on Validation of Analytical Procedures** (1994)
 - <http://www.fda.gov/cder/guidance/ichq2a.pdf>
 - **Q2B: Validation of Analytical Procedures : Methodology** (1996)
 - <http://www.fda.gov/cder/guidance/1320fnl.pdf>
- From Q2B:
 - "For the establishment of linearity, a **minimum of five concentrations is recommended**"
 - "Repeatability should be assessed using (1) a **minimum of 9 determinations covering the specified range for the procedure** (e.g., 3 concentrations/3 replicates each); or (2) a minimum of 6 determinations at 100 percent of the test concentration."

ICH Method Validation Parameters

<http://www.waters.com/waters/division/contentd.asp?watersit=JDRS-SLT6WZ>

Method validation provides an assurance of reliability during normal use, and is sometime referred to as "the process of providing documented evidence that the method does what it is intended to do."

Precision

- "The closeness of agreement between independent test results obtained under stipulated conditions."
- "Precision depends only on the distribution of random errors and does not relate to the true value or specified value. The measure of precision is usually expressed in terms of imprecision and computed as a standard deviation of the test results."
- "A measure for the reproducibility of measurements within a set, that is, of the scatter or dispersion of a set about its central value."

EURACHEM Guide (1998) *The Fitness for Purpose of Analytical Methods: A Laboratory Guide to Method Validation and Related Topics*, p. 45; available at <http://www.eurachem.ul.pt/guides/valid.pdf>

Accuracy

- "The closeness of agreement between a test result and the accepted reference value."
- "Accuracy of a measuring instrument is the ability of a measuring instrument to give responses close to a true value."

EURACHEM Guide (1998) *The Fitness for Purpose of Analytical Methods: A Laboratory Guide to Method Validation and Related Topics*, pp. 39, 41; available at <http://www.eurachem.ul.pt/guides/valid.pdf>

Sensitivity

- **Limit of detection (LOD)** – “the lowest content that can be measured with reasonable statistical certainty.”
- **Limit of quantitative measurement (LOQ)** – “the lowest concentration of an analyte that can be determined with acceptable precision (repeatability) and accuracy under the stated conditions of the test.”
- How low can you go?



EURACHEM Guide (1998) *The Fitness for Purpose of Analytical Methods: A Laboratory Guide to Method Validation and Related Topics*, p. 43; available at <http://www.eurachem.ul.pt/guides/valid.pdf>

Specificity

- “The ability of a method to measure only what it is intended to measure.”
- “Specificity is the ability to assess unequivocally the analyte in the presence of components which may be expected to be present. Typically these might include impurities, degradants, matrix, etc.”
- The primers in PCR amplification provide specificity in forensic DNA testing.

EURACHEM Guide (1998) *The Fitness for Purpose of Analytical Methods: A Laboratory Guide to Method Validation and Related Topics*, p. 51; available at <http://www.eurachem.ul.pt/guides/valid.pdf>

Reproducibility

- “Precision under reproducibility conditions, i.e. conditions where test results are obtained with the same method on identical test items in different laboratories with different operators using different equipment.”
- Will you get the same result each time you test a sample?
- Different from **repeatability**, which is the “precision under repeatability conditions, i.e. conditions where independent test results are obtained with the same method on identical test items in the same laboratory by the same operator using the same equipment within short intervals of time.”

EURACHEM Guide (1998) *The Fitness for Purpose of Analytical Methods: A Laboratory Guide to Method Validation and Related Topics*, pp. 47-48; available at <http://www.eurachem.ul.pt/guides/valid.pdf>

Robustness (Ruggedness)

- “The robustness of an analytical procedure is a measure of its capacity to remain unaffected by small, but deliberate variations in method parameters and provides an indication of its reliability during normal usage.”
- The method works routinely...
- You do not want the method to fail when you only have enough material for a single try.

EURACHEM Guide (1998) *The Fitness for Purpose of Analytical Methods: A Laboratory Guide to Method Validation and Related Topics*, p. 49; available at <http://www.eurachem.ul.pt/guides/valid.pdf>

The lifecycle of a method of analysis



Feinberg et al. (2004) *Anal. Bioanal. Chem.* 380: 502-514

Useful Papers on Validation

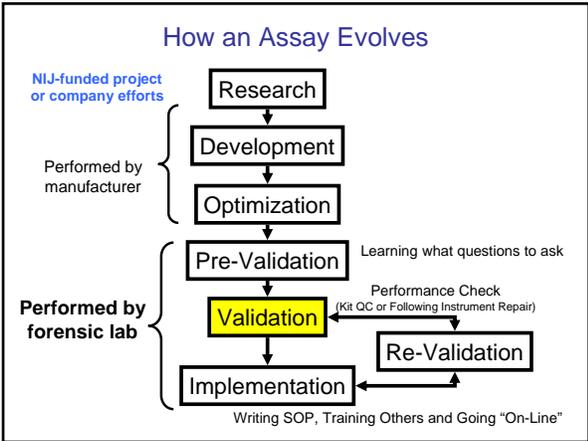
- Taylor JK. (1981) Quality assurance of chemical measurements. *Analytical Chemistry* 53(14): 1588A-1596A.
- Taylor JK. (1983) Validation of analytical methods. *Analytical Chemistry* 55(6): 600A-608A.
- Green JM. (1996) A practical guide to analytical method validation. *Analytical Chemistry* 68: 305A-309A.
- EURACHEM Guide (1998) *The Fitness for Purpose of Analytical Methods: A Laboratory Guide to Method Validation and Related Topics*; available at <http://www.eurachem.ul.pt/guides/valid.pdf>

Developmental Validation

DNA Advisory Board Quality Assurance Standards

Section 2. Definitions

- (ff) Validation is a process by which a procedure is evaluated to determine its **efficacy and reliability** for forensic casework analysis (*DNA analysis*) and includes:
 - (1) Developmental validation is the acquisition of test data and determination of conditions and limitations of a new or novel DNA methodology for use on forensic samples; Manufacturer
 - (2) Internal validation is an accumulation of test data within the laboratory to demonstrate that established methods and procedures perform as expected in the laboratory. Forensic Lab



Validation Section of the DNA Advisory Board Standards

issued October 1, 1998 and April 1999; published in *Forensic Sci. Comm.* July 2000

STANDARD 8.1 The laboratory shall use validated methods and procedures for forensic casework analyses (*DNA analyses*).

8.1.1 Developmental validation that is conducted shall be appropriately documented.

8.1.2 Novel forensic DNA methodologies shall undergo developmental validation to ensure the accuracy, precision and reproducibility of the procedure. The developmental validation shall include the following:

- 8.1.2.1 Documentation exists and is available which defines and characterizes the locus.
- 8.1.2.2 Species specificity, sensitivity, stability and mixture studies are conducted.
- 8.1.2.3 Population distribution data are documented and available.
 - 8.1.2.3.1 The population distribution data would include the allele and genotype distributions for the locus or loci obtained from relevant populations. Where appropriate, databases should be tested for independence expectations.

8.1.3 Internal validation shall be performed and documented by the laboratory.

- 8.1.3.1 The procedure shall be tested using known and non-probative evidence samples (*known samples only*). The laboratory shall monitor and document the reproducibility and precision of the procedure using human DNA control(s).
- 8.1.3.2 The laboratory shall establish and document match criteria based on empirical data.
- 8.1.3.3 Before the introduction of a procedure into forensic casework (*database sample analysis*), the analyst or examination team shall successfully complete a qualifying test.
- 8.1.3.4 Material modifications made to analytical procedures shall be documented and subject to validation testing.

8.1.4 Where methods are not specified, the laboratory shall, wherever possible, select methods that have been published by reputable technical organizations or in relevant scientific texts or journals, or have been appropriately evaluated for a specific or unique application.

FORENSIC SCIENCE COMMUNICATIONS JULY 2000 VOLUME 2 NUMBER 3

Developmental Validation Overview

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PowerPlex Y Developmental Validation Experiments

| Study Completed (17 studies done) | Description of Samples Tested (performed in 7 labs and Promega) | # Run |
|--|---|------------------------------------|
| Single Source (Concordance) | 5 samples x 8 labs | 40 |
| Mixture Ratio (male:female) | 6 labs x 2 M:F mixture series x 11 ratios (1:0, 1:1, 1:10, 1:100, 1:500, 1:1000, 0.5:300, 0.25:300, 0.125:300, 0.0625:300, 0.03:300 ng M:F) | 132 |
| Mixture Ratio (male:male) | 6 labs x 2 M:M mixtures series x 11 ratios (1:0, 19:1, 9:1, 5:1, 2:1, 1:1, 1:2, 1:5, 1:9, 1:19, 0:1) | 132 |
| Sensitivity | 7 labs x 2 series x 6 amounts (1/0.5/0.25/0.125/0.06/0.03) | 84 |
| Non-Human | 24 animals | 24 |
| NIST SRM | 6 components of SRM 2395 | 6 |
| Precision (ABI 3100 and ABI 377) | 10 ladder replicates + 10 sample replicated + [8 ladders + 8 samples for 377] | 36 |
| Non-Probative Cases | 65 cases with 102 samples | 102 |
| Stutter | 412 males used | 412 |
| Peak Height Ratio | N/A (except for DYS385 but no studies were noted) | |
| Cycling Parameters | 5 cycles (28/27/26/25/24) x 8 punch sizes x 2 samples | 80 |
| Annealing Temperature | 5 labs x 5 temperatures (54/58/60/62/64) x 1 sample | 25 |
| Reaction volume | 5 volumes (50/25/15/12.5/6.25) x [5 amounts + 5 concentrations] | 50 |
| Thermal cycler test | 4 models (480/240/960/9700) x 1 sample + [3 models x 3 sets x 12 samples] | 76 |
| Male-specificity | 2 females x 1 titration series (0-500 ng female DNA) x 5 amounts each | 10 |
| TaqGold polymerase titration | 5 amounts (1.382, 0.62, 753, 444, 13 U) x 4 quantities (1/0.5/0.25/0.13 ng DNA) | 20 |
| Primer pair titration | 5 amounts (0.5x/0.75x/1x/1.5x/2x) x 4 quantities (1/0.5/0.25/0.13 ng DNA) | 20 |
| Magnesium titration | 5 amounts (1/1.25/1.5/1.75/2 mM Mg) x 4 quantities (1/0.5/0.25/0.13 ng DNA) | 20 |
| Krenke et al. (2005) <i>Forensic Sci. Int.</i> 148:1-14 | | TOTAL SAMPLES EXAMINED 1269 |

Internal Validation Overview

8.1.3 **Internal validation shall be performed and documented by the laboratory.**

8.1.3.1 The procedure shall be tested using known and non-probative evidence samples (*known samples only*). The laboratory shall monitor and document the reproducibility and precision of the procedure using human DNA control(s).

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| | |
|---|--|
| <h3>Material Modification</h3> <ul style="list-style-type: none">• Decrease in reaction volume from manufacturer's specifications• Centricon tube membrane change• Minimum peak threshold• Injection times for genetic analyzers• Increased amplification cycle numbers• Others? | <h3>Performance Check</h3> <ul style="list-style-type: none">• Relocation of lab to a new facility• Change of laser or other critical component on a genetic analyzer• Software changes<ul style="list-style-type: none">– Mac-based GS/GT to NT-based GS/GT– Mac-based collection software to NT or Windows-based collection software• Additional instrumentation (i.e., 2nd 3130) |
|---|--|

How would you evaluate each of these?

From Robyn Ragsdale (FDLE), Validation Workshop (Aug 24-26, 2005 at NFSTC)
<http://www.cstl.nist.gov/biotech/strbase/validation/validationworkshop.htm>

Other DAB Standards to Consider:

9.1.1 The laboratory shall have an **standard protocol** for each analytical technique used.

9.1.2 The procedures shall include **reagents, sample preparation, extraction, equipment and controls**, which are standard for DNA analysis and data interpretation.

9.2.3 The laboratory shall identify **critical reagents** (if any) and evaluate them prior to use in casework.....

9.4 The laboratory shall monitor the analytical procedures using appropriate **controls and standards**.

10.2 The laboratory shall identify **critical equipment** and shall have a documented program for calibration of instruments and equipment.

10.3 The laboratory shall have a **documented program** to ensure that instruments and equipment are properly maintained.

From Robyn Ragsdale (FDLE), Validation Workshop (Aug 24-26, 2005 at NFSTC)
<http://www.cstl.nist.gov/biotech/strbase/validation/validationworkshop.htm>

Instrument Calibration

STANDARD 10.2 The laboratory (*shall identify critical equipment and*) shall have a documented program for calibration of instruments and equipment.

10.2.1 Where available and appropriate, **standards traceable to national or international standards shall be used for the calibration.**

10.2.1.1 Where traceability to national standards of measurement is not applicable, the laboratory shall provide **satisfactory evidence of correlation of results.**

10.2.2 The frequency of the calibration shall be documented for each instrument requiring calibration. Such documentation shall be retained in accordance with applicable Federal or state law.

Internal Validation

General Steps for Internal Validation

- **Review literature and learn the technique**
- Obtain equipment/reagents, if necessary
- Determine necessary validation studies (there can be overlap and you only need to run a total of 50 samples)
- Collect/obtain samples, if necessary
- **Perform validation studies maintaining all documentation**
- Summarize the studies and submit for approval to Technical Leader
- Write-up the analytical procedure(s). Include quality assurance (controls, standards, critical reagents and equipment) and data interpretation, as applicable
- Determine required training and design training module(s)
- Design qualifying or competency test

From Robyn Ragsdale (FDLE), Validation Workshop (Aug 24-26, 2005 at NFSTC)
<http://www.cstl.nist.gov/biotech/strbase/validation/validationworkshop.htm>

Design of Experiments Conducted for Validation Studies

- Before performing a set of experiments for validation, ask yourself:
 - What is the purpose of the study?
 - Do we already know the answer?
 - Can we write down how we know the answer?
- Think before you blindly perform a study which may have no relevance (e.g., extensive precision studies)
- **Too often we do not differentiate learning, validation, and training**

Points for Consideration

- Remove as many variables as possible in testing an aspect of a procedure
 - e.g., create bulk materials and then aliquot to multiple tubes rather than pipeting separate tubes individually during reproducibility studies
- Who can do (or should do) validation...
 - Outside contractor?
 - Summer intern?
 - Trainee?
 - Qualified DNA analyst

From a validation standpoint, having an outside group perform the validation studies on your instruments is legitimate, **but valuable experience and knowledge are lost...**

Steps Surrounding "Validation" in a Forensic Lab

Effort to Bring a Procedure "On-Line"

This is what takes the time...

- **Installation** – purchase of equipment, ordering supplies, setting up in lab
- **Learning** – efforts made to understand technique and gain experience troubleshooting; can take place through direct experience in the lab or vicariously through the literature or hearing talks at meetings
- **Validation of Analytical Procedure** – tests conducted in one's lab to verify range of reliability and reproducibility for procedure
- **SOP Development** – creating interpretation guidelines based on lab experience
- **QC of Materials** – performance check of newly received reagents
- **Training** – passing information on to others in the lab
- **Qualifying Test** – demonstrating knowledge of procedure enabling start of casework
- **Proficiency Testing** – verifying that trained analysts are performing procedure properly over time

Revised SWGDAM Validation Guidelines (July 2004)

http://www.fbi.gov/hq/lab/fsc/backissu/july2004/standards/2004_03_standards02.htm

3. Internal Validation
 ...a total of at least 50 samples
 (some studies may not be necessary...)

The document provides validation guidelines and definitions approved by SWGDAM July 10, 2003.

Overview of Internal Validation Studies

3. Internal Validation: The internal validation process should include the studies detailed below encompassing **a total of at least 50 samples**. Some studies may not be necessary due to the method itself.

- 3.1 Known and nonprobative evidence samples
- 3.2 Reproducibility and precision
- 3.3 Match criteria
- 3.4 Sensitivity and stochastic studies
- 3.5 Mixture studies
- 3.6 Contamination
- 3.7 Qualifying test

SWGDAM Revised Validation Guidelines
 http://www.fbi.gov/hq/lab/fsc/backissu/july2004/standards/2004_03_standards02.htm

Practical Examples

Practical Examples

- Profiler Plus/COfiler kit switch to Identifiler
- ABI 3100 upgrade to ABI 3130xl
- GeneScan/Genotyper to GeneMapperID
- New allelic ladder provided by company
- Bringing Quantifiler “on-line” (from Quantiblot)
- DNA IQ
- Corbett robot
- FSS-i3 expert system software
- Reduced volume reactions

Discuss each example - participants to provide what they would do...

Suggestions for an Internal Validation of an STR Kit

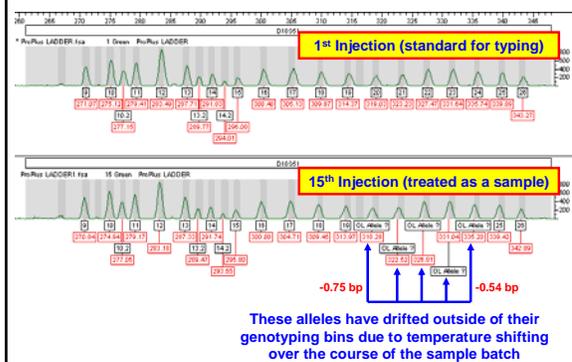
- Standard samples (3.1) **Between 1 and ~20 samples**
 - Verify correct type with positive control or NIST SRM samples
 - Concordance study with 5-10 (non-probative casework) samples previously typed with other kit(s)
- Precision samples (3.2) **5-10 samples**
 - Run at least 5-10 samples (allelic ladder or positive control)
- Sensitivity samples (3.4) **14 samples**
 - Run at least 2 sets of samples covering the dynamic range
 - 5 ng down to 50 pg—e.g., 5, 2, 1, 0.5, 0.2, 0.1, 0.05 ng
- Mixture samples (3.5) **10 samples**
 - Run at least 2 sets of samples
 - Examine 5 different ratios—e.g., 10:1, 3:1, 1:1, 1:3, 1:10

>50 samples

Additional Suggestions for Meeting the SWGDAM Revised Validation Guidelines

- Match Criteria (3.3)
 - As part of running a batch of samples (e.g., 10 or 96), run one allelic ladder at the beginning and one at the end
 - If all alleles are typed correctly in the second allelic ladder, then the match criteria (i.e., precision window of +/-0.5 bp) has likely been met across the entire size range and duration of the run
- Contamination Check (3.6)
 - Run negative controls (samples containing water instead of DNA) with each batch of PCR products
- Qualifying Test (3.7)
 - Run proficiency test samples

Use of Second Allelic Ladder to Monitor Potential Match Criteria Problems



Example with Identifiler STR Kit

- Your lab is currently running ProfilerPlus/COfiler and wants to switch to Identifiler. What is needed for your internal validation?
- What is different between Identifiler and ProfilerPlus/COfiler?
 - Two new STR loci: D19S433 and D2S1338
 - Different fluorescent dyes
 - Additional fluorescent dye (5-dye vs 4-dye)
 - Different dye on internal size standard
 - More loci being amplified in the multiplex
 - Mobility modifiers to move allele sizes
- PCR primer sequences are the same so potential allele discordance due to primer binding site mutations should not be an issue
- What has been reported in terms of developmental validation for Identifiler?

Different
 Loci (2 extra STRs)
 Dyes
 Mobility Modifiers
 Software (5-dye)

ABI Kit Validation Papers

J. Forensic Sci. 2002; 47(1): 66-96

Cydne L. Holt,¹ Ph.D.; Martin Buonocristiani,² M.P.H.; Jeanette M. Wallin,¹ M.P.H.; Theresa Nguyen,¹ B.S.; Katherine D. Lazaruk,¹ Ph.D.; and P. S. Walsh,¹ M.P.H.

TWGDAM Validation of AmpF/STR™ PCR Amplification Kits for Forensic DNA Casework

J. Forensic Sci. 2004; 49(6): 1265-1277

Patrick J. Collins,¹ B.A.; Lori K. Hennessy,¹ Ph.D.; Craig S. Leibelt,¹ A.B.; Rhonda K. Roby,^{1,3} M.P.H.; Dennis J. Reeder,¹ Ph.D.; and Paul A. Foxall,¹ Ph.D.

Developmental Validation of a Single-Tube Amplification of the 13 CODIS STR Loci, D2S1338, D19S433, and Amelogenin: The AmpF/STR® Identifiler® PCR Amplification Kit

AmpF/STR® Identifiler™
 PCR Amplification Kit
 User's Manual

Example: PowerPlex 16

- Switch from ProfilerPlus/COfiler kits to PowerPlex 16
- Retaining same instrument platform of ABI 310

Recommendations:

- Concordance study (somewhat, but better to review literature to see impact across a larger number of samples and which loci would be expected to exhibit allele dropout-e.g., D5S818)
- Stutter quantities, heterozygote peak height ratio
- Some sensitivity studies and mixture ratios
- **Do not need precision studies to evaluate instrument reproducibility**

Example: ABI 3130

- Evaluation of a new ABI 3130 when a laboratory already has experience with ABI 310
- STR kits used in lab will remain the same

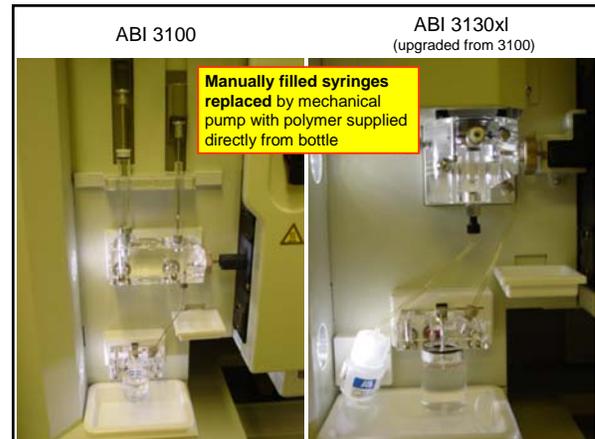
Recommendations:

- Precision studies to evaluate instrument reproducibility
- Sensitivity studies
- **Do not need new stutter, mixture ratio, peak height ratio, etc. (these relate to dynamics of the the kit used)**

Instrument/Software Upgrades or Modifications

- What should be done to “validate” new upgrade?
 - ABI 7000 to ABI 7500
 - ABI 3100 to ABI 3130xl
 - GeneScan/Genotyper to GeneMapper/ID
- Try to understand what is different with the new instrument or software program compared to the one you are currently using (e.g., ask other labs who may have made the switch)
- If possible, try to retain your current configuration for comparison purposes for the validation period

Run the same plate of samples on the original instrument/software and the new one



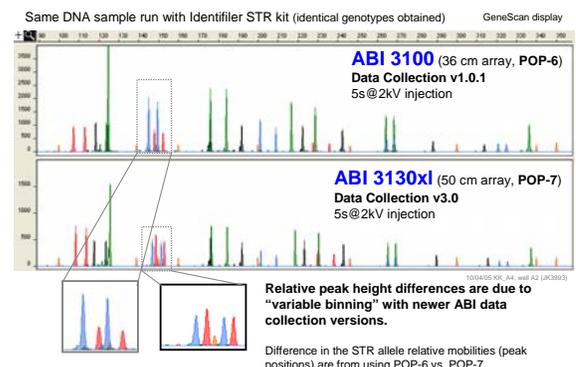
ABI 3130xl vs ABI 3100

What NIST did to “validate” a 3130xl upgrade

- Ran plates of samples on both instruments with same injection and separation parameters and compared results
 - Data Collection version 1.0.1 (3100) vs 3.0 (3130xl)
 - POP-6 (3100) vs POP-7 (3130xl)
 - 36 cm array (3100) vs 50 or 80 cm array (3130xl)
- Ran several plates of Identifier samples and compared allele calls (noticed a sensitivity difference with equal injections and relative peak height differences between dye colors) – **all obtained allele calls were concordant**
- Ran a plate of Profiler Plus samples and compared sizing precision – **precision was not significantly different**
- Also examined SNaPshot products and mtDNA sequencing data

Environmental conditions may change over time so original validation is no longer valid...

Comparison of ABI 3100 Data Collection Versions



Documentation

Documentation of Internal Validation Studies

What is the best way to do this? Standardized format?

Who needs to review?

Who needs to approve?

Should it be presented or published?

From Robyn Ragsdale (FDLE), Validation Workshop (Aug 24-26, 2005 at NFSTC)
<http://www.cstl.nist.gov/biotech/strbase/validation/validationworkshop.htm>

Appropriate Documentation...

- Publications in the Peer-Reviewed Literature
 - See provided reference list
 - <http://www.cstl.nist.gov/biotech/strbase/validation.htm>
- In terms of documentation, is the community doing too much? Too little?
 - Benefit of STRBase Validation website
- Should we be requesting more information from the manufacturers of commercial kits in terms of developmental validation studies?

ABI 7500 Quantifiler Validation Documentation

<http://www.appliedbiosystems.com>



Experimental data supports that the 7500 system with v1.2.3 software provides consistent performance when compared to the ABI PRISM® 7000 Sequence Detection System previously validated for forensic applications. Therefore, the 7500 system can be sold to Human Identification customers at this time. Further guidance for specific operating conditions will follow.

Validation of the Applied Biosystems 7500 Real-Time PCR System with v1.2.3 Software

Applied Biosystems scientists have conducted experiments following the guidance provided by the DAB/SWGDM to validate the Applied Biosystems 7500 Real-Time PCR System with v1.2.3 software ("7500 System") for use in forensic applications using the Quantifiler Human and Quantifiler Y kits. We are pleased to let you know that the Applied Biosystems 7500 Real-Time PCR System equipped with v1.2.3 software is now validated for use in forensic sample testing pursuant to these guidelines using the Quantifiler kits. We conducted experiments, reviewed data, and determined that the 7500 Real-Time PCR System provides results that are robust, reliable, reproducible and provide accurate results when used in conjunction with the Quantifiler™ Human and Quantifiler™ Y kits for the analysis of genomic DNA samples.

Promega Material Modification Reported for PP16 Primer Mix Storage

<http://www.promega.com/applications/hmnd/11072-AN-G1-final.pdf>



Amplifications Using the PowerPlex® 16 System and a 10X Primer Pair Mix Stored in TE⁻ Buffer or in Water Yield Comparable Results

Abstract
Promega is changing the solution used to prepare the 10X Primer Pair Mix in the PowerPlex® 16 System from water to TE⁻ buffer. We show that the amplification results with the 10X Primer Pair Mix prepared in TE⁻ buffer are comparable to those with the 10X Primer Pair Mix prepared in water, even when we varied reaction parameters like amount of DNA template, volume of TE⁻ buffer, the thermal cycle used, amount of AmpliTaq Gold® DNA polymerase used and primer concentrations. Because the performance with both primer pair formulations is comparable, users of the PowerPlex® 16 System should not see adverse effects on amplification results.

Introduction
DNA is traditionally stored in water or a buffered solution, such as TE⁻ (10mM Tris-HCl, 10mM EDTA [pH 8.0]). DNA stored in TE⁻ buffer is more stable due to the buffering capacity of Tris and the presence of EDTA (1). For this reason, the 10X Primer Pair Mix for the

DNA polymerase and 1.5mM MgCl₂, unless otherwise indicated. DNA templates were diluted in 19.2ul of TE⁻ buffer, unless otherwise indicated. Amplifications were carried out using the Perkin-Elmer GeneAmp® PCR System 9600, unless otherwise indicated, for 32 cycles (10/22 cycles). Amplification products were detected using the ABI PRISM® 310 or 3100 Genetic Analyzer and the GeneScan® analysis software. Sample files were imported into the Genotyper® program and analyzed using the PowerType™ in Macro.

Variations in Amplification Reaction Conditions
Primers stored in TE⁻ buffer are more stable than primers stored in water (data not shown), but we wanted to be sure that the use of TE⁻ buffer to resuspend the primers did not affect amplification results. We varied the amplification conditions and compared the results obtained with the two primer pair formulations to determine if there were any effects due to the new formulation.

Validation Section of the DNA Advisory Board Standards issued July 1998 (and April 1999); published in Forensic Sci. Comm. July 2000

STANDARD 8.1

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8.1.1 Developmental validation that is conducted shall be appropriately documented.

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Why is Documentation of Validation Important?

9. Documentation of Validated Methods

9.1 Once the validation process is complete it is important to document the procedures so that the method can be clearly and unambiguously implemented. There are a number of reasons for this. **The various assessments of the method made during the validation process assume that, in use, the method will be used in the same way each time.** If it is not, then the actual performance of the method will not correspond to the performance predicted by the validation data. Thus the **documentation must limit the scope for introducing accidental variation to the method.** In addition, proper documentation is necessary for auditing and evaluation purposes and may also be required for contractual or regulatory purposes.

9.2 Appropriate documentation of the method will help to ensure that **application of the method from one occasion to the next is consistent.**

EURACHEM Guide (1998) *The Fitness for Purpose of Analytical Methods: A Laboratory Guide to Method Validation and Related Topics*, p. 37; available at <http://www.eurachem.ul.pt/guides/valid.pdf>

New Validation Homepage on STRBase

<http://www.cstl.nist.gov/biotech/strbase/validation.htm>

Validation Information to Aid Forensic DNA Laboratories

Validation Summary Sheets

We are initiating an effort to catalog and disseminate information on validation studies conducted by forensic DNA laboratories. The purpose of this effort is to provide a central repository of information on validation studies conducted by forensic DNA laboratories. The information is being collected and documented in a format that is consistent with the SWGDAM Revised Validation Guidelines and summarized.

Below is listed a compilation of reference bibliographic information for specific Validation Summary Sheets.

| Kit, Assay, or Instrument | Reference | How? |
|---------------------------|------------------|------|
| PowerPlex Y | NIST | How? |
| Profiler Plus | Frank et al. (2) | |
| CE-STR | Li et al. (1) | |
| AMPFSTR Blue | Wright | |
| AMPFSTR Green I | Wright | |

Other information and conclusions

Validation Summary Sheet for PowerPlex Y

| Study Completed (17 studies done) | Description of Samples Tested (performed in 7 labs and Promega) | # Run |
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| Single Source (Concordance) | 5 samples x 8 labs | 40 |
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| Sensitivity | 7 labs x 2 series x 6 amounts (10.0, 5.0, 2.5, 1.25, 0.625, 0.3125) | 84 |
| Non-Human | 24 animals | 24 |
| NIST SRM | 6 components of SRM 2395 | 6 |
| Precision (ABI 3100 and ABI 377) | 10 ladder replicates + 10 sample replicated + 8 ladders + 8 samples for 377 | 36 |
| Non-Probative Cases | 65 cases with 102 samples | 102 |
| Slutter | 412 males used | 412 |
| Peak Height Ratio | N/A (except for DYS385 but no studies were noted) | |
| Cycling Parameters | 5 samples (28/27/26/25/24) x 8 punch sizes x 2 samples | 80 |
| Annealing Temperature | 5 labs x 5 temperatures (54/58/60/62/64) x 1 sample | 25 |
| Reaction volume | 5 volumes (50/25/15/12.5/6.25) x 5 concentrations | 50 |
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| Primer pair titration | 5 amounts (0.5x/0.75x/1x/1.5x/2x) x 4 quantities (10.0, 5.0, 2.5, 1.3 ng DNA) | 20 |
| Magnesium titration | 5 amounts (1.1/1.25/1.5/1.75/2 mM Mg) x 4 quantities (10.0, 5.0, 2.5, 1.3 ng DNA) | 20 |

Krenke et al. (2005) *Forensic Sci. Int.* 148:1-14 TOTAL SAMPLES EXAMINED 1269

Laboratory Internal Validation Summaries

We invite updates to this table. Please contact John Butler john.butler@nist.gov if you would like to add a summary of your laboratory's validation studies with a particular forensic DNA test, instrument, or software program. Please submit information in a standard format summarizing the studies conducted, a description of samples run, and the number of samples examined using this downloadable Excel file [\[click here\]](#).

| Kit, Assay, or Instrument | Laboratory | Submitter |
|-------------------------------|---|---------------------|
| PowerPlex 16 Kit with ABI 310 | Pennsylvania State Police | Christina Tomary |
| Quantifiler with ABI 7000 | Alabama Department of Forensic Sciences | Angelo Drulis Manna |

Soliciting Information on Studies Performed by the Community

We can benefit from cumulative experience in the field rather than just single lab results...

| Sample Name | Sample Type | Quantifiler | Quantifiler Result | Percent Difference (NIST - QF)/(QF*100) |
|-------------|-----------------|-------------|--------------------|---|
| SRM1620-10 | negative male | 0.000 | 0.12 | 24.00 |
| SRM1620-11 | positive female | 0.000 | 0.12 | 24.00 |
| SRM1620-12 | positive female | 0.000 | 0.12 | 24.00 |
| SRM1620-13 | negative male | 0.000 | 0.12 | 24.00 |
| SRM1620-14 | positive female | 0.000 | 0.12 | 24.00 |
| SRM1620-15 | positive female | 0.000 | 0.12 | 24.00 |
| SRM1620-16 | positive female | 0.000 | 0.12 | 24.00 |

Example of Validation Documentation

Alabama Department of Forensic Sciences
 Birmingham DNA

ABI Prism® 7000 Validation



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The validation studies referenced above have been reviewed and provide the necessary documentation required by the FBI Director's "Quality Assurance Standards for Forensic DNA Testing Laboratories" for a quantitative method to be used in the forensic casework sections of the Alabama Department of Forensic Sciences Birmingham DNA Laboratory.

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Date

Available on STRBase Validation Website:
http://www.cstl.nist.gov/biotech/strbase/validation/ADFS-BH_7000val.pdf

Documentation of Alabama Validation for ABI 7000 and Quantifiler Assay

What Section of QAS Validation Requirements

Experiments Performed

Summary of Results

Conclusions

8.1.3.1(e) Has the procedure been tested using known and non-probative evidence samples?

Experiment: Sixteen (16) evidence samples composed of various origins that are represented in routine casework were analyzed with the Quantifiler Human Kit on the ABI 7000. These quantitative results were then compared to the previously obtained Quantifiler results. All samples were first amplified using the Identifier Kit and the Quantifiler Human results as an effort to determine the preferred amount of sample template to add to the PCR process.

Results: Additionally, this laboratory participated in a NIST Quantitative study of evaluating changing conditions of standard DNA samples. Each of the NIST samples were analyzed with the Quantifiler Human Kit as well as the Identifier Kit, with results compared and tabulated as well.

| Sample Name | Sample Type | Quantifiler | Quantifiler Result | Percent Difference (NIST - QF)/(QF*100) |
|-------------|-----------------|-------------|--------------------|---|
| SRM1620-10 | negative male | 0.000 | 0.12 | 24.00 |
| SRM1620-11 | positive female | 0.000 | 0.12 | 24.00 |
| SRM1620-12 | positive female | 0.000 | 0.12 | 24.00 |
| SRM1620-13 | negative male | 0.000 | 0.12 | 24.00 |
| SRM1620-14 | positive female | 0.000 | 0.12 | 24.00 |
| SRM1620-15 | positive female | 0.000 | 0.12 | 24.00 |
| SRM1620-16 | positive female | 0.000 | 0.12 | 24.00 |

The experimental results demonstrate that the Quantifiler method of quantitating DNA typically underestimated the amount of DNA present in a sample. An accurate quantitation result is critical to obtaining an adequate DNA profile downstream with the Identifier Kit. If DNA quantities greater than the optimal range are added to the PCR mix, the analyst will likely have a more imbalanced PCR product as well as possible saturation of the detection system causing pull-up and a greater likelihood of stochastic effects. When utilizing the Quantifiler results to determine DNA template addition, the resulting peak heights on the ABI 310 from the Identifier amplicon were acceptable and produced no excessive pull-up or stochastic related issues.

http://www.cstl.nist.gov/biotech/strbase/validation/ADFS-BH_7000val.pdf

Implementation of the Newly Validated Procedure

Ok, the validation studies are complete and approved, the procedure is written and approved and the lab is ready to implement the new procedure into casework.

So, what about training?

Who needs to be trained and what is the extent of the training? How is the training documented? What constitutes completion of training? Per individual or per lab?

From Robyn Ragsdale (FDLE), Validation Workshop (Aug 24-26, 2005 at NFSTC)
<http://www.cstl.nist.gov/biotech/strbase/validation/validationworkshop.htm>

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