

Update from the NIST Applied Genetics group

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April 4, 2018
17th Annual Forensic DNA Conference – Bode 2018



Applied Genetics Group – Forensic & Clinical Genetics












Topics

- NIST Standard Reference Materials (SRMs)
 - 2372a (Human DNA Quantitation), 2391d (PCR-based Profiling)
- STRBase 2.0
- Rapid DNA Study
- Sequencing projects

SRM 2372a - Human DNA Quantitation Standard

- On sale **March 26, 2018**
- https://www-s.nist.gov/srmors/view_detail.cfm?srn=2372a
- Certified by dPCR measurements



Table 1. Certified Values of Number and Mass Concentration for SRM 2372aSM
The copy number values are metrologically traceable to the natural units count 1 and ratio 1 and International System of Units (SI) derived units of volume. The DNA mass concentration values are metrologically traceable to the natural units count and ratio 1 and SI derived units of mass and volume.

Component	Copy Number SM (per µL)	DNA SM (ng/µL)
A (red cap)	15.1 ± 1.5	49.8 ± 5.0
B (white cap)	17.5 ± 1.8	57.8 ± 5.8
C (blue cap)	14.5 ± 1.5	47.9 ± 4.8

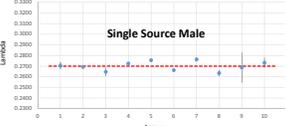
To be used as a qPCR calibrant
OR to assign a value to in house or commercial DNAs

Quantification by dPCR a method to count DNA copies



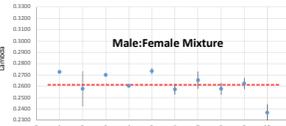
Values assigned by 10 human custom genomic assays Probing single copy targets in the human genome

Component A



Single Source Male

Component C



Male:Female Mixture

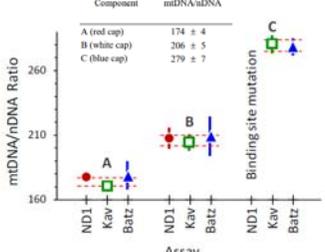
All samples behave as expected with commercial qPCR assays (11 tested)

SRM 2372a includes the ratio of mitochondrial to nuclear haploid genomes

Component	mtDNA/nDNA
A (red cap)	174 ± 4
B (white cap)	206 ± 5
C (blue cap)	279 ± 7

mtDNA/nDNA ratio for three mitochondrial quantification assays optimized for dPCR.

SRM 2372a provides the ratio of mtDNA to gDNA, which bridges the gap between well characterized mtDNA quantification assays and availability of a commercial standard.



Candidate SRM 2391d



- Successor to SRM 2391c
- Similar format – six tubes
 - A-C three single source components
 - D one mixture; approximately 3:1 (F:M)
 - E-F two components: cells spotted on FTA paper (from cell lines)
- Components A-D are DNA extracted from blood (not cell lines)
- Certified allele calls for core STR loci
- Characterized by CE- and NGS-based methods

Data Collection for Sample Screening: mtDNA

Illumina mtDNA Whole Genome Sequencing protocol with Nextera XT Sample Prep Kit

EMPOP results:
https://empop.online/haplotypes#matches_details

Haplogroup	Ancestry	Match
L1b1a12	African	unique

STRBase 2.0

Short Tandem Repeat DNA Internet Database (STRBase)

Introduction:
 Being the forensic DNA and human identity testing community for 20 years, these data are intended to benefit research and application of short tandem repeat DNA markers to human identity testing. The authors are solely responsible for the information herein.

While the use of STRs for genetic mapping and identity testing has become widespread among DNA typing laboratories, there has been a single place where information may be found regarding STR systems. This web site is intended to bring together the distributed literature on the subject for a collective resource to make future work on STRs easier. Aims and objectives of this site are to provide a comprehensive, up-to-date, and accessible resource for STR systems, PCR primer and conditions, and a collection of forensic methodologies for analysis of STR alleles. Data have been included from academic, scientific, and organizational working in this area have also been included as well as a comprehensive reference listing of material on STRs used for DNA typing purposes.

Software Developed by:
 Curtis Miller, Stephen Lee and Maria Norkko

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STRBase 2.0

- First round of development
 - STR fact sheets (for 24 loci)
 - Variant allele reporting
 - Goal is to have a beta site up this summer
- Provide search, sort, and download functionalities
- Automated submission of variant alleles
- Embedded viewer for STR sequence and presentations
- Other ideas? – let us know

STR Fact sheet: example D1S1656

Visualization of the Sequence

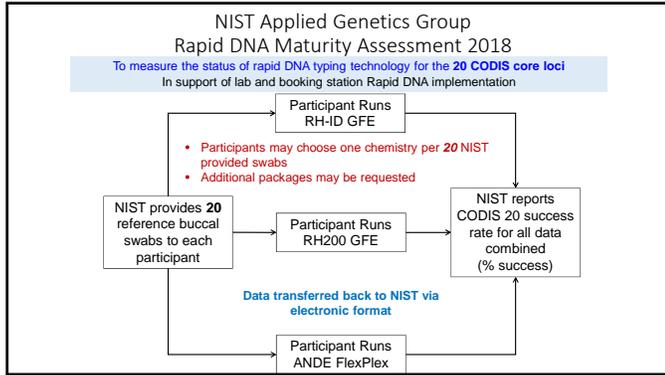
Identify surrounding sequence and provide observed SNPs

Viewer will link to NCBI Bioproject

Rapid DNA Assessment III

- Spring 2018
- Core 20 STR markers
- Projected: 8 labs and 2 vendors
- ANDE and IXI platforms (new kits, configurations)
- 20 samples per lab (single source swab)
- Currently collecting single source swabs

Profiles from high quality single source samples generated by Rapid DNA instruments



- ### Requirements for Participation
- #### Participants

 - Contact Erica Romsos of desired participation in the 2nd Rapid DNA Maturity Assessment
 - Establish Material Transfer Agreement (MTA) with NIST for transfer of samples to each participating lab
 - This can take place while samples are being prepared
 - Participants are responsible for purchasing kits/cartridges
 - Chemistry **must** contain 20 CODIS Core Loci

NIST

 - Collect and characterize buccal reference samples
 - Shipment of samples to all participants
 - Collection, retention, analysis of data from all participants
 - Through a NIST provided FTP site
 - Reporting success metrics to Rapid DNA community
- Questions? Contact Erica Romsos (NIST): erica.romsos@nist.gov**

- ### Sequencing Projects
-
- FGx and S5 platforms
 - **1036 population samples**
 - **Highly polymorphic locus SE33**
 - **STRSeq resource**
 - **Nomenclature support**
 - Sensitivity studies
 - Concordance projects

- ### Applications for Sequencing STRs
- Targeted sequencing of STRs
 - STR motif sequence variation; flanking region variation
 - Further understand simple versus complex repeat motifs
 - Characterize stutter
 - Greater degree of multiplexing
 - Not confined by dye colors; smaller PCR amplicons (degraded samples)
 - PCR for sample enrichment
 - **Still using PCR** – stochastic effects, stutter

- Applications
 - One to one matching?
 - With the new U.S. core loci we are already quite high (>10⁻²⁰)
 - Partial profiles
 - Kinship
 - Mixtures
 - Resolve alleles identical by length, but differ by sequence
 - Separate stutter from low level contributors (based on sequence)
 - A sequenced allele *may* have a lower frequency (higher RMP or LR)
- Allele frequencies of sequenced STR alleles are needed to formally apply this gain in information -> **Generate population data!**

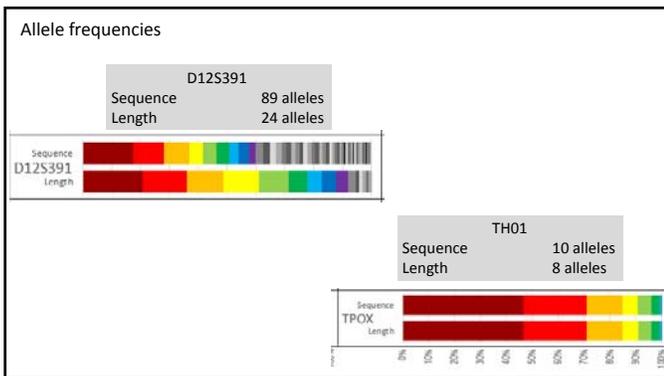
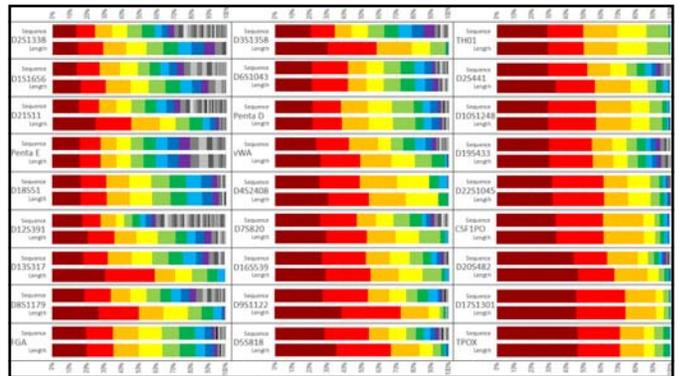
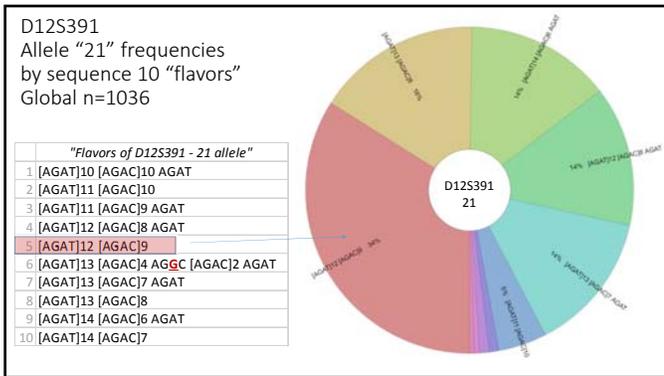
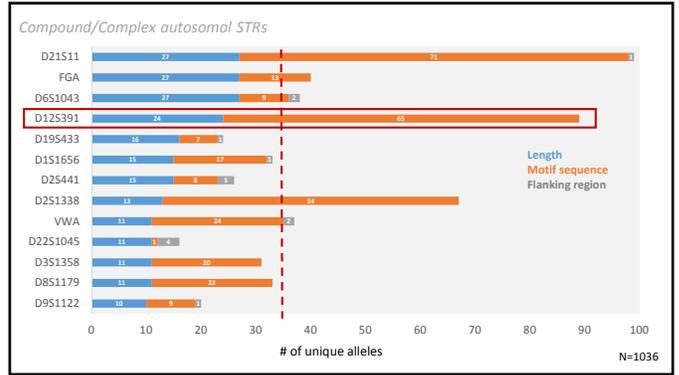
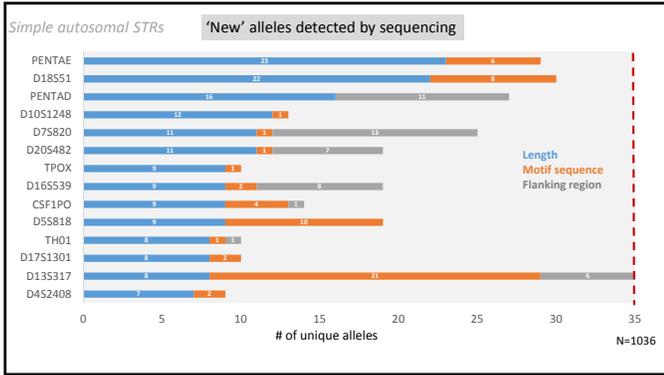
Sequencing Forensic STRs in Population Samples

When a match is made in a forensic case, allele frequencies are used to calculate how common or rare the DNA profile is in a given population

Example of **length** versus **sequence**-based frequency calculation:

D4S2408						Length		Sequence	
Allele	N	Freq	Sequence Allele	N	Freq				
7	1	0.6%	[ATCT]7	1	0.6%	8,9	[ATCT]8, [ATCT]9		
8	23	14.4%	[ATCT]8	23	14.4%	2pq	2pq		
9	60	37.5%	[ATCT]9	18	11.3%	2*0.144*0.375	2*0.144*0.113		
			[ATCT]GTCT [ATCT]7	42	26.3%				
10	53	33.1%	[ATCT]10	53	33.1%	0.108	0.033		
11	21	13.1%	[ATCT]11	21	13.1%				
12	2	1.3%	[ATCT]12	2	1.3%	1 in 9.3	1 in 30.7		

- ### Sequencing of 1036 NIST population samples
- Work performed on Illumina FGx – ForenSeq kit
 - Allele calls were made with Illumina-UAS and STRait Razor and compared to CE length-based calls (**high confidence**)
 - Will include flanking region variation (SNPs, indels)
 - Purpose: provide sequence allele frequencies for four U.S. Population groups
 - U.S.: Caucasian, African American, Hispanic, Asian
 - The manuscript will be submitted by the end of April
 - Focus on the autosomal loci

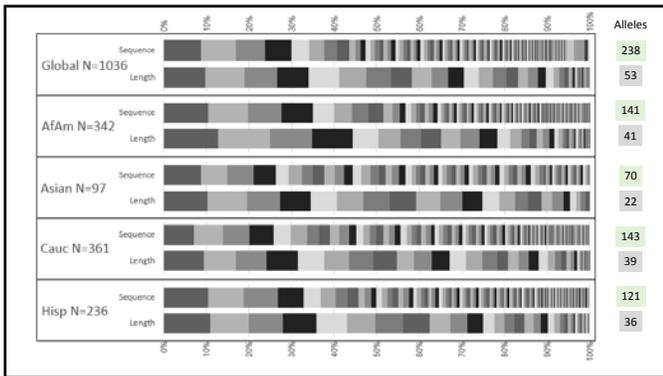


Sequencing of SE33 for 1036 samples

- Separate publication (Electrophoresis Special Issue)
- Focus solely on SE33
- Required further bioinformatic curation of the FGx sequence files

- 53 alleles by length
- 238 allele by sequence
- Data supported by CE-based allele calls

	CE	NGS	CE	NGS
	N	H obs		
All	53	238	0.935	0.965
Cauc	39	143	0.950	0.981
Asian	22	70	0.928	0.959
AfAm	41	141	0.927	0.956
Hispanic	36	121	0.928	0.958



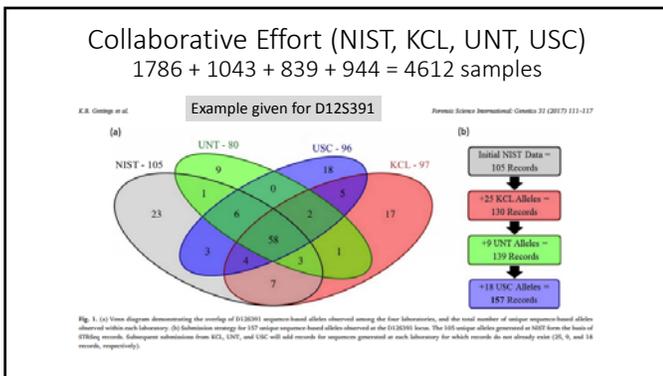
STRSeq

Forensic Science International: Genetics

Research paper
STRSeq: A catalog of sequence diversity at human identification Short Tandem Repeat loci
 Katherine Butler Gettings^{1,2}, Lisa A. Borsak³, David Ballard⁴, Martin Bodner⁵, Bruce Budowle^{6,7}, Laurence Devesse⁸, Jonathan King⁹, Walther Parson¹⁰, Christopher Phillips¹¹, Peter M. Vallone¹²

- Bioproject hosted at NCBI to catalog *unique* STR alleles
 - <https://www.ncbi.nlm.nih.gov/bioproject/380127>
- Fully annotated sequence of the STR amplicon
- Each STR allele will have an accession number

<https://strider.online/>



STRSeq

- A unique record for each observed allele
- CE-based allele call
- Genomic coordinates
- Platform/kit used to generate sequence

Bioproject hosted at NCBI to catalog

- <https://www.ncbi.nlm.nih.gov/bioproject/380127>

Fully annotated sequence of the STR amplicon

- Each STR allele will have an accession number

Nomenclature Support

Forensic Science International: Genetics

STR allele sequence variation: Current knowledge and future issues

Massively parallel sequencing of forensic STRs: Considerations of the DNA commission of the International Society for Forensic Genetics (ISFG) on minimal nomenclature requirements

Defining annotated STR reference sequences

<https://strider.online>

"The devil's in the detail": Release of an expanded, enhanced and dynamically revised forensic STR Sequence Guide

Thank you for your attention! Questions?

Contact: Peter.Vallone@nist.gov

APPLIED GENETICS

- Funding
 - NIST Special Programs Office: *Forensic DNA*
 - FBI Biometrics Center of Excellence: *Forensic DNA Typing as a Biometric tool.*
 - NIJ: *STRSeq and Nomenclature*
 - DHS S&T: *Rapid DNA for Kinship*
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