

**Mixture Interpretation:  
An Open Discussion on Issues and  
Ways to Increase Casework Throughput**

**John M. Butler**  
NIST Biotechnology Division

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**NIST and NIJ Disclaimers**

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<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

**Presentation Format**

**Opening Remarks** (~15-20 minutes):

- Mixtures: what are they and why are they challenging?
- Review of NIST mixture studies and lessons learned
- Observations from MIX05 Interlaboratory Study

**Open Discussion** (~15-20 minutes):

- **Questions and Responses** from audience

**Summary** (~5 minutes):

- Tools available and planned to aid mixture interpretation

**Mixtures: Issues and Challenges**

From J.M. Butler (2005) *Forensic DNA Typing*, 2<sup>nd</sup> Edition, p. 154

- Mixtures arise when two or more individuals contribute to the sample being tested.
- Mixtures can be challenging to detect and interpret without extensive experience and careful training.
- Differential extraction can help distinguish male and female components of many sexual assault mixtures.

**Mixtures: Issues and Challenges**

From J.M. Butler (2005) *Forensic DNA Typing*, 2<sup>nd</sup> Edition, p. 155

- The probability that a mixture will be detected improves with the use of more loci and genetic markers that have a high incidence of heterozygotes.
- The detectability of multiple DNA sources in a single sample relates to the ratio of DNA present from each source, the specific combinations of genotypes, and the total amount of DNA amplified.
- Some mixtures will not be as easily detectable as other mixtures.

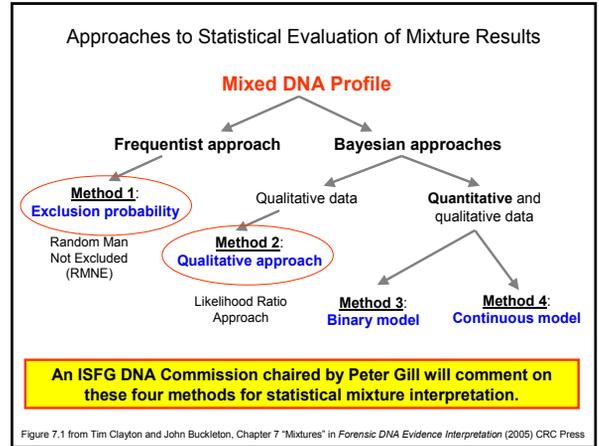
**When is a Sample a Potential Mixture?**

According to several MIX05 participant interpretation guidelines

- Number of Observed Peaks
  - Greater than two peaks at a locus
  - More than two alleles are present at two or more loci, although three banded patterns can occur
  - Presence of 3 alleles at a single locus within a profile
  - 4 peaked patterns (if observed at any locus), 3 peaked patterns (if observed at two or more loci), significant imbalances (peak height ratios <60%) of alleles for a heterozygous genotype at two or more loci with the exception of low template amplifications, which should be interpreted with caution
- Imbalance of heterozygote alleles
  - thresholds range from 50-70%
- Stutter above expected levels
  - generally 15-20%

### Two Parts to Mixture Interpretation

- **Deduction of alleles present in the evidence** (compared to victim and suspect profiles)
- **Providing some kind of statistical answer** regarding the weight of the evidence
  - An ISFG DNA Commission (Peter Gill, Bruce Weir, Charles Brenner, etc.) is evaluating the statistical approaches to mixture interpretation and will make recommendations soon



### Two Parts to Mixture Interpretation

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### NIST Initiated Interlaboratory Studies

Studies involving STRs	# Labs	Publications
Evaluation of CSF1PO, TPOX, and TH01	34	Kline MC, Duewer DL, Newall P, Redman JW, Reeder DJ, Richard M. (1997) Interlaboratory evaluation of STR triplex CTT. <i>J. Forensic Sci.</i> 42: 897-906
Mixed Stain Studies #1 and #2 (Apr–Nov 1997 and Jan–May 1999)	45	Duewer DL, Kline MC, Redman JW, Newall P, J. Reeder DJ. (2001) NIST Mixed Stain Studies #1 and #2: interlaboratory comparison of DNA quantification practice and short tandem repeat multiplex performance with multiple-source samples. <i>J. Forensic Sci.</i> 46: 1199-1210
Mixed Stain Study #3 (Oct 2000–May 2001)	74	Kline, M.C., Duewer, D.L., Redman, J.W., Butler, J.M. (2003) NIST mixed stain study 3: DNA quantitation accuracy and its influence on short tandem repeat multiplex signal intensity. <i>Anal. Chem.</i> 75: 2463-2469. Duewer, D.L., Kline, M.C., Redman, J.W., Butler, J.M. (2004) NIST Mixed Stain Study #3: signal intensity balance in commercial short tandem repeat multiplexes. <i>Anal. Chem.</i> 76: 6928-6934.
DNA Quantitation Study (Jan–Mar 2004)	80	Kline, M.C., Duewer, D.L., Redman, J.W., Butler, J.M. (2005) Results from the NIST 2004 DNA Quantitation Study. <i>J. Forensic Sci.</i> 50(3):571-578
Mixture Interpretation Study (Jan - Aug 2005)	69	<b>MIX05</b> <i>Data analysis currently on-going ...</i>

### Overall Lessons Learned from NIST MSS 1,2,&3

- Laboratories have instruments with different sensitivities
- Different levels of experience and training plays a part in effective mixture interpretation
- Amount of input DNA makes a difference in the ability to detect the minor component (labs that put in "too much" DNA actually detected minor components more frequently)

### Mixture Interpretation Interlab Study (MIX05)

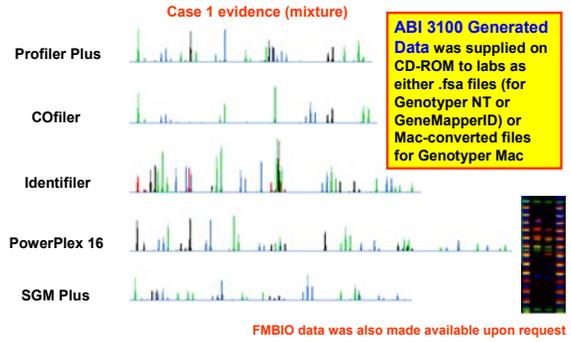
- **Only involves interpretation of data – to remove instrument detection variability and quantitation accuracy issues**
- **94 labs enrolled** for participation
- **69 labs have returned results** (17 from outside U.S.)
- Four mock cases supplied with "victim" and "evidence" electropherograms (GeneScan .fsa files – that can be converted for Mac or GeneMapper; gel files made available to FMBIO labs)
- Data available with Profiler Plus, COfiler, SGM Plus, PowerPlex 16, Identifier, PowerPlex 16 BIO (FMBIO) kits

### MIX05 Study Design and Purpose

- Permit a large number of forensic practitioners to evaluate the same mixture data
- Provide multiple cases representing a range of mixture scenarios
- Generate data from multiple STR kits on the same mixture samples to compare performance for detecting minor components
- The primary variable should be the laboratory's interpretation guidelines rather than the DNA extraction, PCR amplification, and STR typing instrument sensitivity
- Are there best practices in the field that can be advocated to others?

### MIX05 Results on Multiple Kits

<http://www.cstl.nist.gov/biotech/strbase/interlab/MIX05.htm>



### Requests for Participants in MIX05

Mixtures representing four different case scenarios have been generated at NIST with multiple STR kits and provided to laboratories as electropherograms.

We would like to receive the following information:

- 1) Report the results as though they were from a real case including whether a statistical value would be attached to the results. Please summarize the perpetrator(s) alleles in each "case" as they might be presented in court—along with an appropriate statistic (if warranted by your laboratory standard operating procedure) and the source of the allele frequencies used to make the calculation. Please indicate which kit(s) were used to solve each case.
- 2) Estimate the ratio for samples present in the evidence mixture and how this estimate was determined.
- 3) Provide a copy of your laboratory mixture interpretation guidelines and a brief explanation as to why conclusions were reached in each scenario

### Summary of MIX05 Responses

94 labs enrolled for participation  
69 labs returned results (17 from outside U.S.)

50 labs made allele calls  
39 labs estimated ratios  
29 labs provided stats

**STR kit results used**  
34 ProfilerPlus/COfiler  
10 PowerPlex 16  
7 PP16 BIO  
5 Identifier  
2 SGM Plus  
1 All ABI kit data  
9 Various combinations

### Summary of Some MIX05 Reported Results

Case #2 Yuse Peep	2779619	D1S1581	VWA	FGA	AMEL	D18S1179	D17S11	D16S11	D15S10	D13S17	D7S820	D16S539	TH01	TPOX	CSF1PO
LabID	Kit Used	15, 15	15, 15, 20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10	
18	ProfilerPlus/COfiler	--	--	--	--	--	--	--	--	--	--	--	--	--	--
6	ProfilerPlus/COfiler	15	15	20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
91	SGM Plus	15	15	20, 24	X,Y	11, 13	20, 32, 2	17, 18				10, 11	7, 9, 3		
48	PP16	--	--	--	--	--	--	--	--	--	--	--	--	--	--
37	ProfilerPlus/COfiler	--	15	20	X,Y	13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
2	PP16	15	15, 15, 20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	INC	8, 10	8, 10	10, 11	7, 9, 3	9, 10	7, 10
13	PP16 & Identifier	15	15	20, 24	--	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
34	ProfilerPlus/COfiler	15	15	20, 24	--	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
70	Identifier	15	15	20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
55	ProfilerPlus/COfiler	15	15	20, 24	--	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
21	ProfilerPlus/COfiler	15, 15	15, 15, 20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10	
73	ProfilerPlus/COfiler	15, 15	15, 15, 20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10	
29	ProfilerPlus/COfiler	15, 15	15, 15, 20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10	
71	ProfilerPlus/COfiler	15, 15	15, 15, 20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10	
23	ProfilerPlus/COfiler	15, 15	15, 15, 20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10	
29	Identifier	15	15	20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
54	All kits	15, 15	15, 15, 20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10	
90	ProfilerPlus/COfiler	15	15	20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
9	ProfilerPlus/COfiler	15	15	20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
4	ProfilerPlus/COfiler	15	15	20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
33	ProfilerPlus/COfiler	--	--	--	--	--	--	--	--	--	--	--	--	--	--
12	ProfilerPlus/COfiler	15	15	20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
67	PP16	15	15, 15, 20, 24	--	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10	
86	ProfilerPlus/COfiler	15, 15	15, 15, 20, 24	--	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10	
77	Identifier	--	--	--	--	--	--	--	--	--	--	--	--	--	--
69	PP16	15	15	20, 24	X,Y	11, 13	20, 32, 2	17, 18	8, 13	12, 14	8, 10	10, 11	7, 9, 3	9, 10	7, 10
61	Identifier	--	--	--	--	--	--	--	--	--	--	--	--	--	--

Most calls were correct (when they were made)

### Some Mixture Ratios Reported in MIX05

Many labs do not routinely report the estimated ratio of mixture components

LabID	Case1 (F:M)	Case2 (M:F)	Case3 (M:F)	Case4 (F:M)
13	2	5	<2	10
34	1.8-3.6	3.9-6.7	1.6-1.8	6.2-7.6
70				
55	68%:32%	85%:15%	64%:36%	
21				
73	2:1	6:1	2:1	not determined
29				
54	2:1	6:1	2:1	6:1
90	male23-39%	not determined	male64-71%	
9	3 or 4:1	4 or 5:1	1.4:1	~10:1
4	10:1	6:1	1:1	not determined
33	male60-78%	male80-90%	male58-71%	victim86%
12	male25%	male85%	male40-45%	unknown10%
67	1.2:3	6.4:1	2:1	1:6.8
86	2:1	6.6:5:1	1.6-2:1	4.4:5:1
79	~3:1 to ~2:1	~6:1 to ~4:1	~2:1*	a lot of victim
77				
60	2:1	5:1	2:1	10:1
61				

Some Reported Stats for MIX05 Case #1

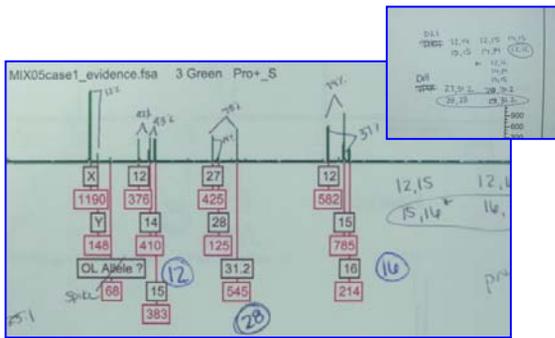
LabID	Kits Used Identifier	Case1		
		Caucasians PE calculated	African Americans PE calculated	Hispanics PE calculated
73	ProPlus/Cofiler	none provided	none provided	none provided
4	ProPlus/Cofiler	none provided	none provided	none provided
12	ProPlus/Cofiler	none provided	none provided	none provided
29	Identifier	none provided	none provided	none provided
90	ProPlus/Cofiler	1.18E+15	2.13E+14	3.09E+15
34	ProPlus/Cofiler	2.40E+11	7.00E+09	9.80E+10
46	PP16	5.60E+09	3.80E+11	none provided
33	ProPlus/Cofiler	2.94E+08	1.12E+08	1.74E+09
6	ProPlus/Cofiler	40,000,000	3,500,000	280,000,000
9	ProPlus/Cofiler	1.14E+07	1.97E+07	1.54E+08
61	Identifier	1.50E+06	260,000	2.40E+07
79	ProPlus/Cofiler	930,000	47,900	1,350,000
16	ProPlus/Cofiler	434,600	31,710	399,100

Some Differences in Reporting Statistics

LabID	Kits Used	Case1		
		Caucasians	African Americans	Hispanics
90	ProPlus/Cofiler	1.18E+15	2.13E+14	3.09E+15
34	ProPlus/Cofiler	2.40E+11	7.00E+09	9.80E+10
33	ProPlus/Cofiler	2.94E+08	1.12E+08	1.74E+09
6	ProPlus/Cofiler	40,000,000	3,500,000	280,000,000
9	ProPlus/Cofiler	1.14E+07	1.97E+07	1.54E+08
79	ProPlus/Cofiler	930,000	47,900	1,350,000
16	ProPlus/Cofiler	434,600	31,710	399,100

Remember that these labs are interpreting the same MIX05 electropherograms

Manual Solving of MIX05 Peak Ratios and Possible Mixture Combinations



Manual Solving of Mixture Component Profiles

Locus	Allele	Peak height	Possible Component profiles giving rise to observed mixture	Comments
D8	12	51.3	12, 15	50.3 / 100.0 = 50.3%
	15	244	12, 15	12:15 not balanced; has no other contributing contribution
D16	29	237	27, 28	if secondary only 2 contributors
	28	487	27, 28	237-272 = 60; 487-282 = 205; 60/205 = 29.3%
	27	155	27, 28	155-272 = -117; 155-282 = -127; 155-144 = 111; 111/144 = 77% balance
D18	12	207	12, 14	if 12:14, 2 pk balance = 70%
	14	331	12, 14	if 12:14, 2 no major
	17	443	14, 17	443-17 = 426; 207-14 = 193; 426/193 = 2.21; 443/331 = 1.34; 443/214 = 2.07
11	387	11, 11	if homozygous, 2 major	

Another MIX05 Participant Manually Solving A Mixture

D21S11	13	1081	1081/1545 = 0.700	1081/1545 = 0.700
D21S11	14	132	132/1545 = 0.085	132/1545 = 0.085
D21S11	28	972	972/1545 = 0.623	972/1545 = 0.623
D21S11	30	164	164/1545 = 0.106	164/1545 = 0.106
D21S11	31	88	88/1545 = 0.057	88/1545 = 0.057
D21S11	22.2	1010	1010/1545 = 0.654	1010/1545 = 0.654
D18S11	12	162	162/300 = 0.540	162/300 = 0.540
D18S11	16	138	138/300 = 0.460	138/300 = 0.460
D18S11	17	96	96/300 = 0.320	96/300 = 0.320
D18S11	18	1033	1033/300 = 3.443	1033/300 = 3.443
D8S11	8	1060	1060/1060 = 1.000	1060/1060 = 1.000
D8S11	11	140	140/1060 = 0.132	140/1060 = 0.132
D8S11	12	232	232/1060 = 0.219	232/1060 = 0.219
D8S11	13	843	843/1060 = 0.795	843/1060 = 0.795
D13S17	8	129	129/270 = 0.478	129/270 = 0.478
D13S17	9	141	141/270 = 0.522	141/270 = 0.522
D13S17	13	905	905/270 = 3.352	905/270 = 3.352
D13S17	14	817	817/270 = 3.026	817/270 = 3.026
D7S820	8	887	887/1018 = 0.871	887/1018 = 0.871
D7S820	9	155	155/1018 = 0.152	155/1018 = 0.152
D7S820	10	805	805/1018 = 0.791	805/1018 = 0.791
D7S820	11	98	98/1018 = 0.096	98/1018 = 0.096
D3S1358	10	1543	1543/1543 = 1.000	1543/1543 = 1.000
D3S1358	16	124	124/1543 = 0.080	124/1543 = 0.080
D18S51	9	282	282/191 = 1.477	282/191 = 1.477
D18S51	10	420	420/191 = 2.199	420/191 = 2.199
D18S51	11	1337	1337/191 = 7.000	1337/191 = 7.000
D18S51	12	213	213/191 = 1.115	213/191 = 1.115
TH01	7	738	738/100 = 7.380	738/100 = 7.380
TH01	8	87	87/100 = 0.870	87/100 = 0.870
TH01	8.5	680	680/100 = 6.800	680/100 = 6.800
TH01	10	81	81/100 = 0.810	81/100 = 0.810

Semi-Automated Locus-by-Locus Interpretation Performed by One MIX05 Participant

Peak Allele	A B C D				Known type: K1 K2	
	28	30	31	32.2	30	31
RFU	988	167	92	1025		
Possible combinations						
28	30	AND	31	32.2	N	N
28	31	AND	30	32.2	N	N
28	32.2	AND	30	31	N	N
Thresholds 70% 60% Htzryg. Mixture ratio(1/2)						
96.39% 55.09%						
Best fit						

Peak Allele	A B C D				Known type: K1 K2	
	12	15	17	18	12	15
RFU	163	138	972	1047		
Possible combinations						
12	15	AND	17	18	Y	Y
12	17	AND	15	18	N	N
12	18	AND	15	17	N	N
Thresholds 70% 60% Htzryg. Mixture ratio(1/2)						
85.28% 92.84% 0.15 Known present						

Excel spreadsheet used to examine possible component combinations



A High Degree of Variability Currently Exists with Mixture Interpretation

- “If you show 10 colleagues a mixture, you will probably end up with 10 different answers”  
– Peter Gill, Human Identification E-Symposium, April 14, 2005
- Is this a problem?

Question

- Without a suspect, should a mixture be considered (by show of hands, Y or N)?



Question

- Can mixture reports be standardized (to make them easier for review)?

Question

- Do you look at the evidence data first without considering the suspect's profile?

Question

- Do you have a decision point whereby you consider a mixture too complicated and do not try to solve it?
- If so, is the case declared inconclusive?

Question

- What elements would be needed in an expert system in order to automatically evaluate mixtures?

Question

- Are composite profiles acceptable – e.g., high injection for minor component and low injection for major component allele identification?

Question

- Should two amplifications be done – e.g., one at 1 ng to type the major component and one at higher concentration to move the minor component out of the low-copy number regime?

Question

- What kind of training materials would be beneficial to help your laboratory more effectively solve mixtures?

Question

- What are the biggest obstacles you face in your lab in terms of mixture interpretation?

Question

- How do you report mixture statistics in court?

Question

- What percentage of time is spent in a case trying to deduce the mixture components?

### Question

- Do more loci in a multiplex result in less minor component alleles detected?...
- Is there an optimal multiplex for mixtures?



### Question

- Will improved training information and software tools aid in mixture interpretation (or will lab policies prevent examination of these cases no matter what tools are brought to bear on this problem)?

## WRAP-UP AND SUMMARY

### Some Final Thoughts...

- It is of the highest importance in the art of detection to be able to recognize out of a number of facts, which are incidental and which vital. Otherwise your energy and attention must be dissipated instead of being concentrated (Sherlock Holmes, *The Reigate Puzzle*).
- **“Don’t do mixture interpretation unless you have to”** (Peter Gill, Forensic Science Service, 1998).

### Purpose of MIX05 Study

- **Goal is to understand the “lay of the land” regarding mixture analysis across the DNA typing community**
- One of the primary benefits we hope to gain from this study is **recommendations for a more uniform approach to mixture interpretation** and training tools to help educate the community

### Value of the MIX05 Study

<http://www.cstl.nist.gov/biotech/strbase/interlab/MIX05.htm>

- Data sets exist with multiple mixture scenarios and a variety of STR kits that **can be used for training purposes**
- A wide variety of approaches to mixture interpretation have been applied on the **same data sets evaluated as part of a single study**
- **Interpretation guidelines from many laboratories are being compared to one another for the first time in an effort to determine challenges facing future efforts to develop “expert systems” for automated mixture interpretation**
- **We are exploring the challenges of supplying a common data set to a number of forensic laboratories** (e.g., if a standard reference data set was ever desired for evaluating expert systems)

### Additional Thoughts on Mixtures

From J.M. Butler (2005) *Forensic DNA Typing*, 2<sup>nd</sup> Edition, p. 166

- Some forensic DNA laboratories may decide not to go through the trouble of fully deciphering the genotype possibilities and assigning them to the major and minor contributors.
- An easier approach is to simply include or exclude a suspect's DNA profile from the crime scene mixture profile. If all of the alleles from a suspect's DNA profile are represented in the crime scene mixture, then the suspect cannot be excluded as contributing to the crime scene stain.
- Likewise, the alleles in a victim's DNA profile could be subtracted out of the mixture profile to simplify the alleles that need to be present in the perpetrator's DNA profile.

### NIST Software Programs to Aid Mixture Work

*Excel-based programs developed by David Duewer (NIST)*

- **mixSTR** (developed at request of Palm Beach Sheriff's Office)
  - Does not interpret data (relies on user inputted alleles following STR data review)
  - Aids in the organization of STR mixture information
  - Considers only the presence/absence of alleles (no peak heights used)
- **Virtual MixtureMaker** (developed to aid MIX05 sample selection)
  - Creates mixture combinations through pairwise comparisons of input STR profiles
  - Returns information on the number of loci possessing 0,1,2,3,4,5, or 6 alleles in each 2-person mixture (also reports number of loci in each sample with 0,1,2, or 3 alleles)
  - Useful for selection of samples in mixture or validation studies with various degrees of overlapping alleles in combined STR profiles
  - Useful in checking for potentially related individuals in a population database

Programs can be downloaded from NIST STRBase web site:  
<http://www.cstl.nist.gov/div831/strbase/software.htm>

### Conclusions

- We plan to develop training information based on lessons learned from the MIX05 study.
- We intend to create other useful software tools like **mixSTR** and **Virtual MixtureMaker** to increase mixture interpretation capabilities of the forensic DNA typing community.

### Acknowledgments

Funding from interagency agreement 2003-IJ-R-029 between NIJ and the NIST Office of Law Enforcement Standards



John Butler   Margaret Kline   Pete Vallone   Mike Coble   Jan Redman   Amy Decker   Becky Hill   Chris DeAngelis   Dave Duewer

#### Role in MIX05

- Margaret Kline (sample prep, running study)
- John Butler (study design and data review)
- Becky Hill (GeneMapper/D data review)
- Jan Redman (Access database entry, shipping)
- Dave Duewer (*Virtual MixtureMaker* to aid sample selection; **mixSTR** program)
- Chris Tomsey & Frank Krist (FMBIO Mac data)
- Kermit Channel & Mary Robnett (FMBIO NT data)

The many forensic scientists and their supervisors who took time out of their busy schedules to examine the MIX05 data provided as part of this interlaboratory study

Thank you for your attention...

Our publications and presentations are available at:  
<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

Contact information:  
[john.butler@nist.gov](mailto:john.butler@nist.gov)  
301-975-4049

<http://www.cstl.nist.gov/biotech/strbase>