

## An Evaluation of CODIS and Non-CODIS MiniSTR Loci

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## Disclaimers

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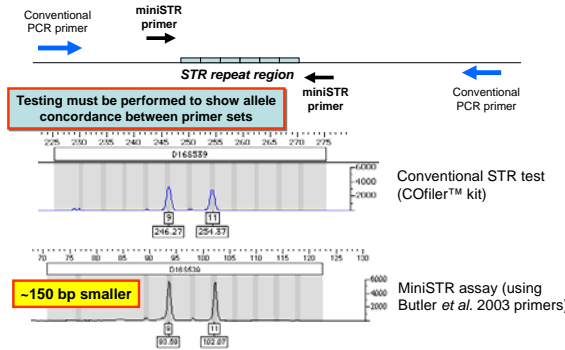
**Our publications and presentations are made available at:**  
<http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm>

## Outline of Topics to Discuss

- Introduction to miniSTR loci
- Concordance study with ABI MiniFiler™ kit (including 7 CODIS loci) using U.S. population samples
- The value of non-CODIS miniSTR loci
- The characterization of additional miniSTR loci
- The “miniMegaplex” 5-dye single amplification multiplex system: *in development...*

## Introduction to MiniSTR Loci

A miniSTR is a reduced size STR amplicon that enables higher recovery of information from degraded DNA samples

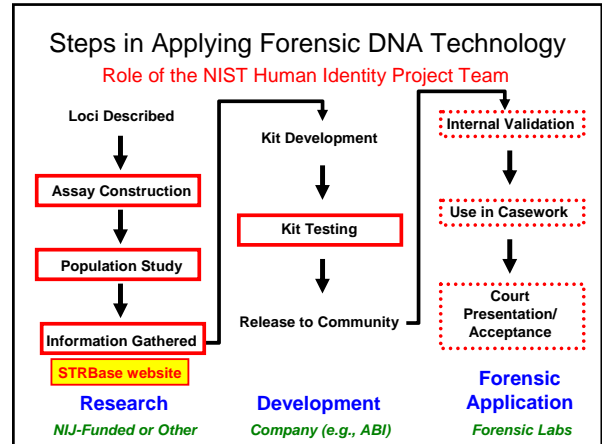


Butler, J.M. (2005) *Forensic DNA Typing, 2nd Edition*, Figure 7.2, ©Elsevier Science/Academic Press

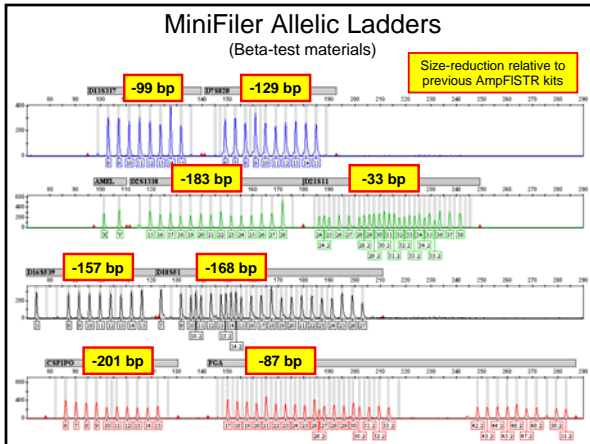
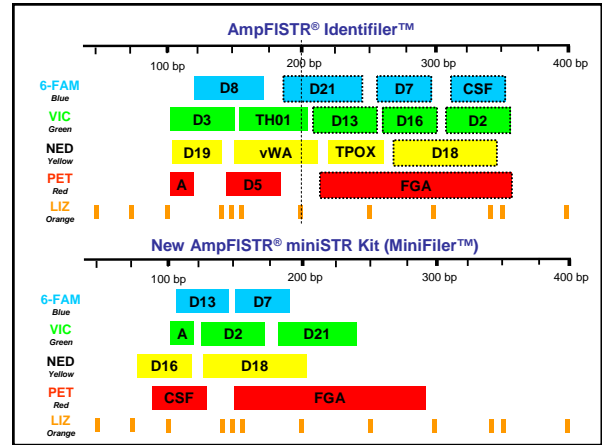
## Timeline for miniSTRs and Demonstrating the Value of Using Reduced Size Amplicons for Degraded DNA

- 1994 – FSS finds that smaller STR loci work best with burned bone and tissue from Branch Davidian fire
- NIJ** 1997 – New primers developed for time-of-flight mass spectrometry to make small STR amplicons
- NIJ** 2001 – Work at NIST and OhioU with CODIS STRs; **BodePlexes used in WTC investigation starting 2002**
- NIJ** 2004 – Work at NIST with **non-CODIS (NC) miniSTRs**
- 2007 – Applied Biosystems releases 9plex MiniFiler  
<http://www.cstl.nist.gov/biotech/strbase/miniSTR/timeline.htm>

How Do We See Our  
Role at NIST?



Concordance Study with ABI  
MiniFiler kit



### Summary of Samples Typed with ABI MiniFiler kit at NIST and ABI

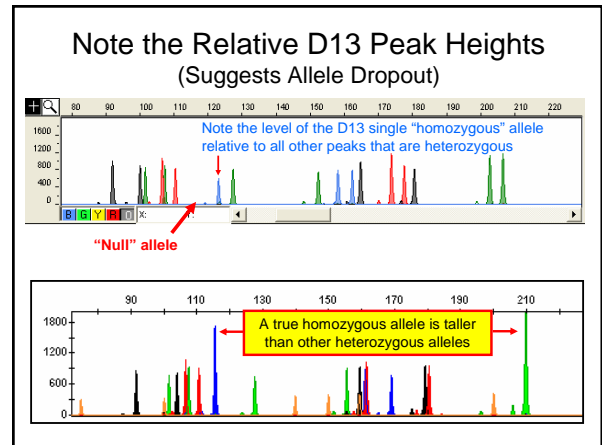
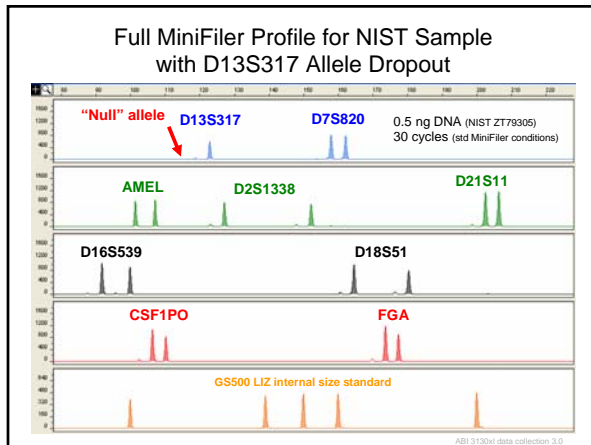
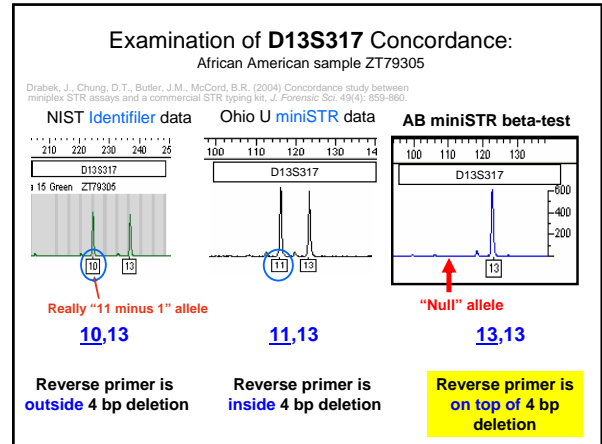
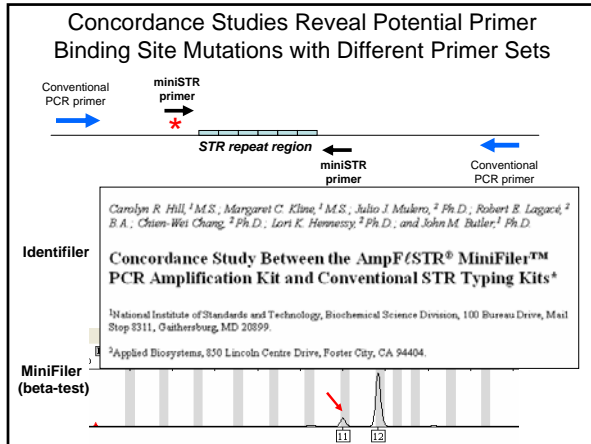
Applied Biosystems  
NIST

- Primarily only population samples examined – no extensive sensitivity or degraded DNA tests were performed

**1,308 samples** Allele concordance = 10,437/10,464 = 99.7%

- 656 NIST U.S. population samples**
  - 260 Caucasian, 253 African American, 140 Hispanic, 3 Asian
  - Previously examined with **Identifiler**; also with **PowerPlex 16**
  - Also tested with Butler *et al.* (2003) published **miniSTR primers**
  - <http://www.cstl.nist.gov/biotech/strbase/NISTpop.htm>
- 481 father-son pairs**
  - 184 Caucasian, 196 African American, 101 Asian samples (provided by paternity testing company DDC)
  - Previously examined with **Identifiler**
- 171 samples from Applied Biosystems**

Hill, C.R., Kline, M.C., Mulero, J.J., Lagace, R.E., Chang, C.-W., Hennessy, L.K., Butler, J.M. (2007) Concordance study between the AmpFISTR MiniFiler PCR Amplification Kit and conventional STR typing kits. *J. Forensic Sci.*, in press.



### Apparent Null Alleles Observed During Concordance Studies

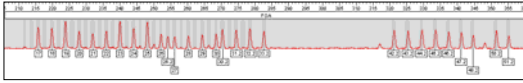
**New Section of STRBase (launched to track MiniFile discordance and allele dropout frequency):**  
<http://www.cstl.nist.gov/biotech/strbase/NullAlleles.htm>

Locus	STR Kits/Assays Compared	Results	Frequency of Primer Binding Site Mutation	Source
CSF1PO	MiniFile vs ID vs F216	MP: 11,11 and ID: 11,11.1 One base insertion in Identifier amplicon outside of MiniFile and F216 primers	1/1308	Hill et al. (2007)
CSF1PO	FF16 vs COfiler	Loss of allele 14 with COfiler, fine with FF16	2/1537	Bradweil et al. (2001)
FGA	SQM vs SQM Plus	Loss of allele 26 with SQM Plus; weak amp of same allele with SQM		Cotton et al. (2000)
FOA	FF16 vs ProFiler	Loss of allele 22 with ProFiler, fine with FF16		Bradweil and Spachner (2001)
TH01	FF16 vs COfiler	Loss of allele 9 with COfiler, fine with FF16	1/1537	Bradweil et al. (2001)
TH01	SQM vs SQM Plus	Loss of allele 6 with SQM Plus, fine with SQM	1/4245	Clayton et al. (2004)
YWA	FF11 vs ProFiler	Loss of allele 19 with ProFiler, fine with FF11	2/1483	Kline et al. (1998) and Walsh (1995)
YWA	FF16 vs ProFiler	Loss of alleles 15 and 17 with ProFiler, fine with FF16	2/1537	Bradweil et al. (2001)
YWA	ID vs multiplex	Loss of alleles 12, 13, and 14 with multiplex assay, fine with ID	9/532	Drabek et al. (2004)

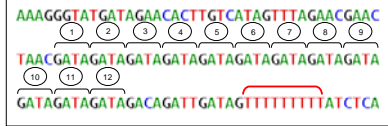
### The Value of Non-CODIS MiniSTR Loci

## Why Go Beyond the CODIS Loci?

(1) Large Allele Ranges (e.g. FGA)

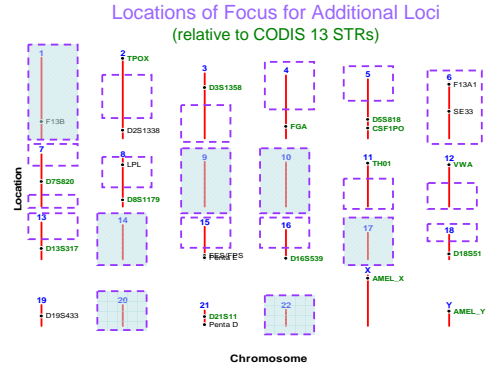


(2) "Unclean" Flanking Sequences (e.g. D7S820)



Butler, JM, Shen, Y., McCord, BR (2003) JFS 48(5): 1054-1064

(3) A large amount of unused chromosomal locations

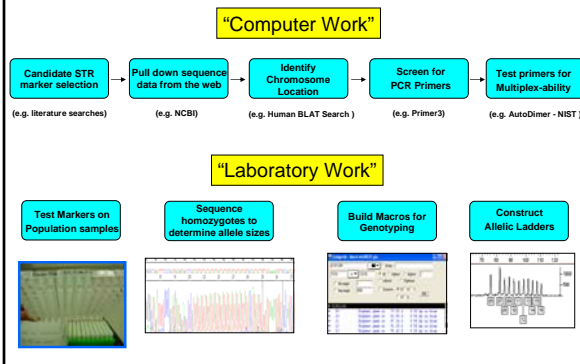


## Uses of Additional Non-CODIS miniSTR Loci in the Forensic Community

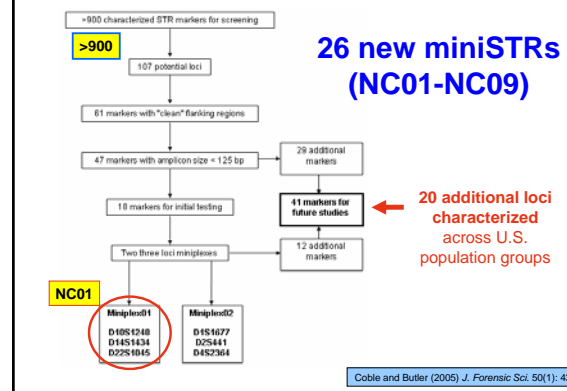
- Obtaining additional information with degraded DNA samples
- Missing persons investigations
- Identification of mass disaster victims
- Paternity testing

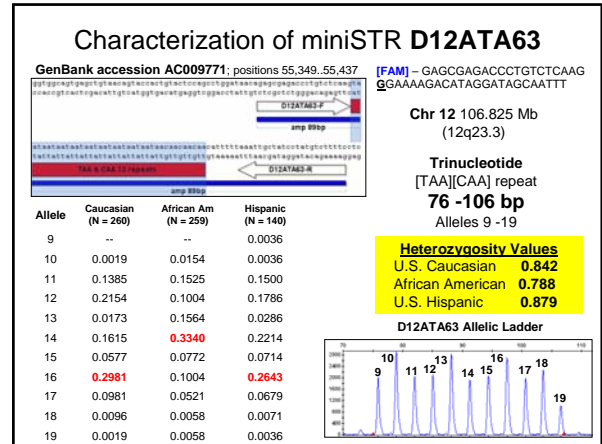
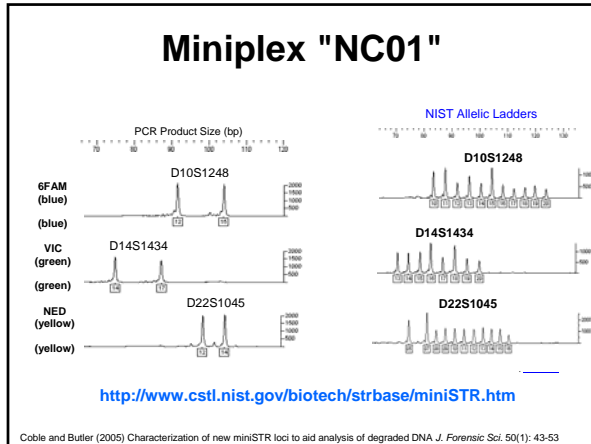
## The Characterization of Additional MiniSTR Loci

## Characterization of New Loci



## Initial Testing Results with Potential miniSTR Loci





### European Labs Have Adopted the NIST-Developed NC (non-CODIS) miniSTRs

*FSI (2006) 156(2): 242-244*

Short communication

The evolution of DNA databases—Recommendations for new European STR loci

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...recommended that existing multiplexes are re-engineered to enable small amplicon detection, and that **three new mini-STR loci with alleles <130 bp (D10S1248, D14S1434 and D22S1045) are adopted as universal. This will increase the number of European standard Interpol loci from 7 to 10.**

(D14 has been replaced with D2S441 from NC02)

Comparison of heterozygosity values on 26 non-CODIS loci across the U.S. samples examined in this study.

Locus	N	Heterozygosity (Overall)	Rank	African American	Caucasian	Hispanic
D9S2157	661	0.844	1	0.884	0.840	0.779
ATA63 (D12)	659	0.829	2	0.788	0.842	0.879
D10S1248 (NC01)	663	0.792	3	0.825	0.785	0.743
D22S1045 (NC01)	663	0.784	4	0.817	0.785	0.721
D2S441 (NC02)	660	0.774	5	0.798	0.780	0.721
D10S1435	663	0.768	6	0.798	0.770	0.700
D2S1776	654	0.763	7	0.740	0.801	0.734
D3S4529	660	0.761	8	0.752	0.723	0.829
D6S474	648	0.761	9	0.765	0.802	0.679
D5S2500	664	0.747	10	0.757	0.747	0.729
D1S1627	660	0.746	11	0.783	0.737	0.693
D1S1677 (NC02)	660	0.746	12	0.743	0.749	0.743
D6S1017	664	0.740	13	0.807	0.698	0.693
D3S3053	648	0.739	14	0.713	0.724	0.814
D9S1122	659	0.734	15	0.753	0.742	0.686
D17S974	664	0.732	16	0.757	0.702	0.743
D11S4463	664	0.730	17	0.780	0.676	0.743
D4S2408	654	0.722	18	0.752	0.709	0.691
D18S853	664	0.711	19	0.772	0.645	0.721
D20S1082	664	0.696	20	0.792	0.653	0.600
D14S1434 (NC01)	663	0.696	21	0.685	0.721	0.650
D20S482	648	0.691	22	0.673	0.689	0.729
GATA113 (D1)	654	0.668	23	0.673	0.632	0.727
D8S1115	664	0.663	24	0.629	0.660	0.729
D17S1301	664	0.649	25	0.626	0.717	0.564
D4S2364 (NC02)	660	0.511	26	0.385	0.551	0.664

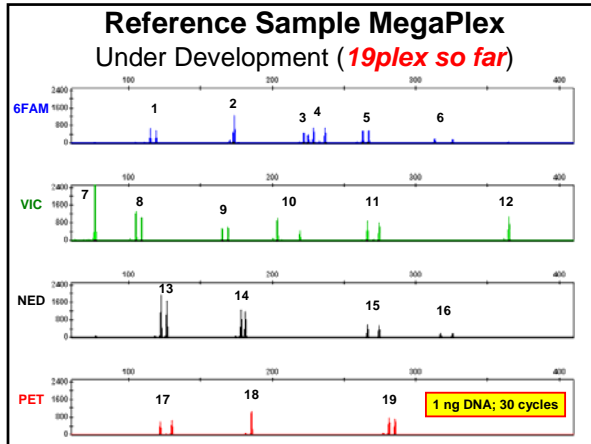
### 26 New miniSTR Loci Typed (and Sequenced) with Standard Samples

Locus	Standard DNA Template Genotypes				SRM 2201b Components							
	9947A	9948	ABI 007	R567	Genomic: 1	Genomic: 2	Genomic: 3	Genomic: 4	Genomic: 5	Genomic: 6	Genomic: 7	Genomic: 8
D15G1A113	11,12	7,12	12,12	11,12	11,11	12,13	11,11	13,13	11,12	11,12	10,12	10,12
D1S1627	13,14	11,13	11,14	10,14	10,14	13,14	13,14	11,12	14,16	11,13	11,14	13,14
D1S1677 (NC02)	13,14	13,14	13,13	13,14	12,13	14,16	14,17	14,15	13,14	13,14	12,13	14,16
D2S441 (NC02)	10,14	11,12	14,15	10,14	11,14	11,14	10,14	12,14	11,14	10,11	11,14	11,13
D2S1776	10,10	10,12	8,10	10,11	11,12	11,11	8,10	11,12	12,13	11,12	11,12	11,12
D3S3053	9,11	9,12	9,9	12,12	9,12	10,11	9,11	11,11	11,11	9,9	11,11	9,9
D3S4529	13,13	12,12	13,13	14,14	14,15	13,16	14,16	15,16	13,15	15,17	14,16	14,14
D4S2364 (NC02)	9,10	9,10	9,10	9,9	9,9	9,10	9,10	9,9	9,10	8,9	9,9	9,9
D4S2408	9,10	10,10	10,11	10,11	10,10	9,9	9,9	9,10	10,11	9,9	8,11	11,11
D5S2500	14,23	14,17	17,18	14,14	17,18	17,24	17,18	17,18	14,16	14,18	14,20	14,18
D6S474	14,18	17,17	14,14	16,18	15,17	14,17	14,15	14,16	15,18	14,17	15,17	17,17
D6S1017	9,10	8,8	10,10	8,11	10,10	10,12	10,12	7,10	8,9	10,10	7,12	10,12
D8S1115	9,18	15,17	15,17	16,16	16,16	16,16	16,17	9,17	9,15	9,16	9,18	15,16
D9S1122	12,13	12,16	12,12	10,14,15	11,12	12,13	12,12	12,12	11,13	11,12	11,12	13,13
D9S1527	7,19	7,19	13,13	13,13	8,13	9,11	11,13	11,11	7,14	11,13	12,16	11,11
D10S1248 (NC01)	13,15	12,16	12,15	12,12	14,16	13,15	13,16	12,12	14,15	14,15	13,14	11,15
D10S1435	10,11	12,13	11,13	10,12	13,13	11,14	13,14	12,12	11,12	12,12	12,12	11,13
D11S4463	12,13	12,14	14,14	13,14	14,14	13,14	14,15	11,12	14,16	16,17	14,15	14,17
D12AT463	13,13	13,18	12,17	17,17	14,17	13,17	12,16	16,18	13,16	14,18	16,17	14,16
D14S1434 (NC01)	11,13	13,14	11,14	10,10	13,14	11,13	14,15	10,11	13,14	13,14	10,14	13,13
D17S974	7,10	10,11	9,10	8,8	9,11	9,10	9,9	7,9	11,12	9,9	11,11	8,9
D17S1301	12,12	11,12	12,13	11,12	11,11	11,12	12,13	11,11	11,11	11,11	11,12	12,12
D18S853	11,14	11,11	11,11	12,16	11,14	11,11	11,11	11,13	10,15	11,14	14,14	12,13
D20S482	14,15	13,14	14,15	15,15	14,14	14,16	15,15	14,15	14,16	14,14	14,14	15,16
D20S1082	11,14	11,15	12,14	11,11	11,15	14,15	11,11	14,15	11,14	11,15	14,15	11,15
D22S1045 (NC01)	11,14	16,18	11,16	16,16	14,15	11,16	15,16	17,18	11,14	11,15	11,15	16,17

[http://www.cstl.nist.gov/biotech/strbase/miniSTR/miniSTR\\_NC\\_loci\\_types.htm](http://www.cstl.nist.gov/biotech/strbase/miniSTR/miniSTR_NC_loci_types.htm)

### "miniMegaplex" 5-dye Single Amplification Multiplex System

In Development...



In Summary

### Conclusions

- **MiniFiler**: concordance study with ABI (Hill *et al. J. Forensic Sci., in press*)
- **New Non-CODIS (NC) Loci: 26 miniSTR loci** tested on NIST **665 U.S. population samples**
- **European forensic community** has recommended three miniSTRs (D2S441, D10S1248, D22S1045) to be adopted as standard core loci
- **SRM 2391b components are being certified** through sequencing for D10S1248, D2S441, D22S1045; for reference purposes, genotypes for standard samples (9947A, 9948, 007, K562) are available on STRBase
- A “**miniMegaplex**” is in development to type 26 miniSTR loci in a 5-dye single reaction
- **New miniSTR loci information on STRBase:**  
<http://www.cstl.nist.gov/biotech/strbase/newSTRs.htm>








### NIST Publications on miniSTR Loci

- Butler, J.M., Shen, Y., McCord, B.R. (2003) The development of reduced size STR amplicons as tools for analysis of degraded DNA. *J. Forensic Sci* 48(5): 1054-1064.
- Coble, M.D. and Butler, J.M. (2005) Characterization of new miniSTR loci to aid analysis of degraded DNA. *J. Forensic Sci.* 50: 43-53.
- Coble, M.D., Hill, C.R., Vallone, P.M., Butler, J.M. (2006) Characterization and performance of new miniSTR loci for typing degraded samples. *Progress in Forensic Genetics 11*, Elsevier Science: Amsterdam, The Netherlands, International Congress Series 1288, 504-506.
- Butler, J.M. (2006) MiniSTRs: past, present, and future. *Forensic News* (Applied Biosystems) October 2006.
- Butler, J.M. and Coble, M.D. (2007) Authors' Response to Letter to Editor [regarding nomenclature for new miniSTR locus D10S1248]. *J. Forensic Sci.* 52(2): 494.
- Hill, C.R., Kline, M.C., Mulero, J.J., Lagace, R.E., Chang, C.-W., Hennessy, L.K., Butler, J.M. (2007) Concordance study between the AmpFISTR MiniFiler PCR Amplification Kit and conventional STR typing kits. *J. Forensic Sci., in press.*
- Hill, C.R., Coble, M.D., Butler, J.M. (2007) Characterization of 26 miniSTR loci for improved analysis of degraded DNA samples. *submitted.*

### Acknowledgments


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**NIST Human Identity Project Team – Leading the Way in Forensic DNA...**

John Butler   Margaret Kline   Pete Vallone   Jan Redman   Amy Decker   Becky Hill   Dave Duewer

Tom Reid (DNA Diagnostics Center) – supplying the father-son samples for use in MiniFiler concordance study



Mike Coble (AFDIL Research Section Chief, former member of NIST Human Identity Team)

Applied Biosystems - Lori Hennessy, Julio Mulero, Robert Legacé, and Chien-Wei Chang